# SCORPION DIVERSITY AND DISTRIBUTION IN SOUTHERN AFRICA: PATTERN AND PROCESS

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#### Abstract:

Patterns of scorpion diversity and distribution in southern Africa (south of 15° latitude), and the processes that produced them, are reviewed. A georeferenced presence-only dataset, comprising 6766 point locality records for the 140 scorpion species currently recognised in the subregion, is compiled and analysed with a geographical information system. Hotspots of scorpion species richness and endemism in southern Africa are mapped at the level of a quarterdegree square. The taxonomic composition of the southern African scorpion fauna is assessed and found to comprise distinct western and eastern components. Hotspots of species richness and endemism are concentrated in arid regions with rugged topography, complex geology, or substratal heterogeneity. The distributions of genera and species are discussed in terms of their ecological requirements and modes of speciation within the context of historical events. Historical changes in the geomorphology and climate of southern Africa, coupled with the specific ecological requirements of most southern African scorpions, are proposed as primary causes for their speciation and, ultimately, their high species richness and endemism.

Key words:

Chelicerata; Scorpiones; southern Africa; diversity; endemism; distribution; GIS; hotspot; taxonomy; evolution; speciation; biogeography

#### 1. INTRODUCTION

Predictable, non-random patterns characterise the spatial distribution of most taxa (MacArthur, 1972; Myers and Giller, 1988), and scorpions are no exception. The distribution of scorpions has interested arthropod biogeographers for more than a century (Pocock, 1894, Kraepelin, 1905;

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Birula, 1917a, 1917b, 1925; Hewitt, 1925; Lawrence, 1952; Lamoral, 1980a; Stockwell, 1989; Sissom, 1990; Nenilin and Fet, 1992; Lourenço, 1996, 1998, 2000; Soleglad and Fet, 2003). Scorpions allow opportunities for ecological and historical biogeographic investigation almost unparalleled among Arthropoda. One reason for using scorpions as model taxa in biogeographic studies is the antiquity of the group. Fossil records show that scorpions have existed in the terrestrial environment since the middle Silurian, over 400 MYA, and are almost unchanged today (Jeram, 1990a, 1990b, 1994a, 1994b; Selden, 1993; Sissom, 1990). Scorpion distribution patterns often reveal evidence of continental drift (Koch, 1977, 1981; Lourenço, 1984a, 1991a, 1996, 1998, 2000; Stockwell, 1989; Sissom, 1990) or palaeoclimatic change (Lourenço, 1994a, 1994b, 1996). A similarity in the habits and ecological requirements of fossil and modern scorpions may be inferred from their morphological conservatism (Jeram, 1990b). It is thus reasonable to assume that the ecological factors determining scorpion distributions today are similar to those which determined their distributions in the past.

Scorpions are also useful for biogeographic studies because of their diversity (Lourenço, 1994c). Scorpions occur on all major landmasses except Antarctica, and on many oceanic islands, in all terrestrial habitats including high-elevation mountaintops, caves, and the intertidal zone except tundra and high-latitude taiga (Polis, 1990; Gromov, 2001). Diverse communities of scorpions (4-13 species) occur in habitats as different as desert (Williams, 1980) and tropical rainforest (González-Sponga, 1978, 1984, 1996; Lourenço, 1983a). Many ecological factors influence the spatial distribution of scorpions, including temperature, precipitation, soil or rock characteristics, stone or litter cover, topography, vegetation, environmental physiognomy (Polis, 1990). Some of these factors, e.g. the substratum, may be important determinants of their evolution and may underpin broad patterns of geographical distribution (Lamoral, 1978a; Prendini, 2001a). For example, the sedentary nature of most scorpion species, together with their often narrow physiological and ecological tolerances (Polis, 1990), may have limited their vagility and promoted allopatric speciation by vicariance during periods of palaeoclimatic change (Lamoral, 1978b; Prendini, 2001a), in turn creating high levels of endemism in certain areas, e.g. Baja California (Williams, 1980).

The prevalence of relictual, endemic scorpion species in certain areas may warrant their biogeographic investigation as models for arthropod conservation (Lourenço, 1991b). The importance of knowing the distribution of venomous scorpions, from a medical perspective, is yet another reason to study their biogeography (Newlands, 1978b; Newlands and Martindale, 1980). The ecological abundance and medical importance of scorpions has also ensured their good representation in museum

collections. The taxonomic diversity of scorpions is thus better understood, worldwide, than that of many other arthropod taxa. Both factors, in turn, guarantee greater accuracy in biogeographic investigations of the distributions of species or higher taxa.

The rich scorpion fauna of the southern African subregion was recognised at the turn of the previous century. Hewitt (1925) identified a difference between the scorpion species composition of the northeastern and southwestern parts of the subregion, and proposed that the most southerly species were primitive, species further north being more advanced. Hewitt's work was published before the theories of continental drift and plate tectonics were widely accepted, and concentrated on dispersalist rather than vicariance arguments. It also failed to address the ecology of the group. Lawrence (1942, 1952) noted the same distinction between the eastern and western scorpion faunas, but attributed the relative paucity of the eastern fauna entirely to ecological factors, while neglecting the role of history. Later theories about scorpion distribution patterns in southern Africa (Newlands, 1972a, 1972b, 1978a, 1978b, 1980; Lamoral, 1978b, 1979) also failed to accommodate ecological explanations within a historical context.

Such criticisms are not reserved solely for studies of scorpion biogeography, but extend to biogeographic studies of many other taxa. Few authors (see, for example, references in Werger (1978) and Keast (1981)) consider patterns of distribution in terms of ecology and history, despite the undeniable influence of both (Haydon et al., 1994). Myers and Giller (1988: 3) warn that "to progress, biogeography must attempt to ... determine how speciation, adaptation, extinction and ecological processes interact with one another and with geology and climate to produce distributional patterns in the world's biota through time."

In this contribution, a distributional dataset for the southern African scorpion species is compiled and analysed with a geographical information system (GIS). Using evidence from the literature, the observed patterns of diversity and distribution are interpreted as the outcome of two interacting sets of circumstances: (1) ecological factors, i.e. the distribution of environmental gradients, the ecological requirements of the species, and the biotic component of species interactions; (2) historical factors, i.e. past, often chance events that have determined the occurrence of a species in particular localities, e.g. dispersal, vicariance, and speciation (Koch, 1977).

#### METHODS

## 2.1 Taxonomic considerations

Traditionally, all southern African scorpions were placed into Buthidae C.L. Koch, 1837 or Scorpionidae Latreille, 1802 (Hewitt, 1918, 1925; Lawrence, 1955; Lamoral and Reynders, 1975; Lamoral, 1978b, 1979; Newlands, 1978a, 1978b, 1980; Newlands and Martindale, 1980; Newlands and Cantrell, 1985). Today, they are divided among Bothriuridae Simon, 1880, Buthidae, Liochelidae Fet and Bechly, 2001, and Scorpionidae.

Bothriuridae are represented in southern Africa by two genera: *Brandbergia* Prendini, 2003; *Lisposoma* Lawrence, 1928. Although previously known to share similarities with the bothriurids (Vachon, 1974), *Lisposoma* was retained in Scorpionidae, in a unique subfamily, Lisposominae Lawrence, 1928 (Lawrence, 1955; Lamoral, 1978b, 1979), until Francke (1982) transferred it to Bothriuridae. Lourenço's (2000) creation of a unique family, Lisposomidae Lawrence, 1928, for the genus is unsupported by cladistic analysis (Prendini, 2000a, 2003a, 2003b). *Lisposoma* was revised by Prendini (2003b).

Six genera of Buthidae were recognised in southern Africa (Buthotus Vachon, 1949; Karasbergia Hewitt, 1913; Lychas C.L. Koch, 1845; Parabuthus Pocock, 1890; Pseudolychas Kraepelin, 1911; Uroplectes Peters, 1861) until a seventh, Afroisometrus Kovařík, 1997, was created to accommodate a species of Lychas from Zimbabwe (FitzPatrick, 1994a). Francke (1985) demonstrated that Buthotus is a junior synonym of Hottentotta Birula, 1908. Southern African Parabuthus have been studied intensively in recent years and 20 species are currently recognised from the subregion (FitzPatrick, 1994b; Prendini 2000b, 2001b, 2003c, 2004a). The species composition of Karasbergia and Pseudolychas remains unaltered following recent revisions (Prendini 2004b, in press). Uroplectes currently contains 19 southern African species (FitzPatrick, 1996, 2001; Fet and Lowe, 2000), but requires extensive revision; the genus is undoubtedly more diverse than Parabuthus.

Liochelidae, until recently known as Ischnuridae Simon, 1879, contains three southern African genera formerly assigned to subfamily Ischnurinae Simon, 1879 of Scorpionidae (Lourenço, 1989; Sissom, 1990): *Cheloctonus* Pocock, 1892; *Hadogenes* Kraepelin, 1894; *Opisthacanthus* Peters, 1861. A fourth genus, *Iomachus* Pocock, 1893, with one species, *Iomachus politus* Pocock, 1896, widespread in eastern Africa (Ethiopia, Kenya, Tanzania, Uganda and the Democratic Republic of Congo), and also recorded from northeastern Mozambique (Kraepelin, 1913; Werner, 1936), has not been confirmed as occurring south of 15° latitude; records of *I. politus* from Beira

(Werner, 1936; Aguiar, 1978) are almost certainly erroneous. *Opisthacanthus* was revised by Lourenço (1987a) but the validity of several southern African species remains questionable, as does the validity of *Cheloctonus* and its component species, which remain to be addressed (Prendini, 2000a). *Hadogenes*, presently containing 16 species, is the subject of ongoing revision by the author (Newlands and Prendini, 1997; Prendini, 2001c, in press). Lourenço's (1999, 2000) proposals to transfer this genus to Scorpionidae, or to create a unique family, Hadogenidae Lourenço, 2000, to accommodate it, are unsupported by cladistic analysis (Prendini, 2000a, 2001c).

Opistophthalmus C.L. Koch, 1837 remains the sole representative of Scorpionidae in southern Africa. This diverse genus, also under revision by the author, currently contains 59 valid species (Prendini, 2000c, 2001d; Harington, 2002). The actual number is closer to 80 (Prendini et al., 2003). To date, no records of Pandinus viatoris (Pocock, 1890), a scorpionid widespread in central Africa (recorded from the Democratic Republic of Congo, Malawi, Tanzania, Zambia, and northwestern Mozambique), have been confirmed as occurring south of 15° latitude. Alleged records of this species from Zimbabwe (Lamoral and Reynders, 1975; Fet, 2000a) are actually in Zambia (Prendini et al., 2003). A single record from Maputo (Aguiar, 1978) is erroneous.

The status of many southern African scorpion species and subspecies, particularly within *Uroplectes* and the liochelid genera, remains contentious in spite of intensive taxonomic work. However, for the purpose of this study, the taxonomy reflected by the most recent published treatments was employed, according to which 140 species are presently recognised from Africa south of 15° latitude (Appendix 1). Subspecies were not considered, although some will certainly be elevated to species in future revisions.

## 2.2 Distributional data

Point locality data for each species were collated from available published locality records. Lamoral and Reynders' (1975) catalogue of the scorpions described from the Afrotropical Region up to December 1973 was used for all the early records. Remaining (post-1975) records were obtained from the following works: Eastwood (1977a, 1977b, 1978a, 1978b); Harington (1978, 1984, 2002); Lamoral (1978b, 1979, 1980b); Lourenço (1981, 1987a); Newlands (1980); Newlands and Martindale (1980); Newlands and Cantrell (1985); Fitzpatrick (1994a, 1994b, 1996, 2001); Newlands and Prendini (1997); Prendini (2000b, 2000c, 2001b, 2001c, 2003a, 2003b, 2003c, 2004a, 2004b, in press).

Literature records were supplemented with records of personally identified specimens deposited in the following collections: Austria: Zoologisches Institut und Naturhistorisches Museum, Universität Wien; Belgium: Musée Royal de l'Afrique Centrale, Tervuren; Denmark: Zoological Museum, University of Copenhagen; France: Museum National d'Histoire Naturelle, Paris; Germany: Forschungsinstitut und Natur-Museum Senckenberg, Frankfurt; Zoologisches Forschungsinstitut und Museum Alexander Koenig, Bonn; Zoologisches Institut und Zoologisches Museum. Universität Hamburg; Zoologisches Museum, Universität Humboldt, Berlin; Namibia: Desert Research Foundation of Namibia, Gobabeb; National Museum of Namibia, Windhoek; South Africa: Albany Museum, Grahamstown; Kruger National Park Reference Collection, Skukuza; KwaZulu-Natal Nature Conservation Service, Pietermaritzburg; McGregor Museum, Kimberley; Natal Museum, Pietermaritzburg; National Collection of Insects and Arachnids, Plant Protection Research Institute, Pretoria; National Museum, Bloemfontein; South African Museum, Cape Town; Transvaal Museum, Pretoria; University of Natal, Pietermaritzburg; Sweden: Göteborgs Naturhistoriska Museet, Göteborg; Naturhistoriska Riksmuseet, Stockholm; Zoologiska Institutionen, Lunds Universitet; Switzerland: Musée d'Histoire Naturelle, La-Chaux-de-Fond; Muséum d'Histoire Naturelle, Genève; The Netherlands: Zoologisch Museum, Universiteit van Amsterdam; U.S.A.: American Museum of Natural History, New York; California Academy of Sciences, San Francisco; Field Museum of Natural History, Chicago; Museum of Comparative Zoology, Harvard University, Cambridge, MA; U.S. National Museum of Natural History, Smithsonian Institution, Washington, DC; U.K.: The Natural History Museum, London; Zimbabwe: Natural History Museum of Zimbabwe, Bulawayo.

## 2.3 Georeferencing

All records of sufficient accuracy were collated to create a point locality geographical dataset for mapping distributional ranges. Only a small proportion of the records were initially accompanied by geographical coordinates or quarter-degree squares (QDS). Localities defined in degrees and minutes latitude and longitude (accurate to within 1.7 km²) were used preferentially, whenever possible, owing to their high level of spatial resolution. All such localities were, in turn, converted to decimal-degree format for GIS input. Many localities were, however, only available in QDS notation (de Meillon et al., 1961). A decimal-degree approximation for the centroid of the QDS was used for these records, as a centroid would always be situated nearest to the average locality. For example, a locality registered

as occurring in QDS SE1927Ac would be recorded as -19.375/27.125. As a QDS covers an area of approximately 26 km² in southern Africa (Poynton, 1964), this may result in some loss of resolution. Augmenting accurate locality records with QDS centroids yields distribution maps at least as accurate as maps using the QDS notation system (Newlands, 1980; Newlands and Martindale, 1980).

Available georeferences were checked for accuracy and an attempt was made to trace coordinates for as many remaining records as possible by reference to gazetteers, the official 1:250 000 and 1:500 000 topo-cadastral maps of Namibia, Botswana, Lesotho, South Africa and Swaziland published by the Government Printer of South Africa, and the GEOnet Names Server: http://164.214.2.59/gns/html/cntry files.html. This database is based on gazetteers published by U.S. National Imagery and Mapping Agency for the U.S. Board on Geographic Names, which contain the standard names approved for official use, unapproved variant names, designations (cities, mountains, rivers, etc.) and coordinates: http://geonames.usgs.gov/bgngaz.html. The following regional gazetteers and lists of place-names were consulted: Gross (1920); Doidge (1950); Webb (1950); National Place Names Committee (1951, 1978, 1988); Ministry of Land and Natural Resources (1959); Gonçalves (1962); Davis & Misonne (1964); Poynton (1964); Copeland (1966); Ministry of Land and Mines (1967); SWALU (1967); Postmaster-General (1970); Surveyor-General (1970, 1988); Coaton and Sheasby (1972); Skead (1973); Haacke (1975); Penrith (1975, 1977, 1979, 1981a,b, 1987); Leistner and Morris (1976); Lamoral (1979); Bamps (1982); Raffle (1984); Poynton and Broadley (1985a,b, 1987); Raper (1987); Irish (1988); Polhill (1988); Flora Zambesiaca: Gazetteer of Localities (unpublished document, Bolus Herbarium Library, University of Cape Town); Herpetology Department, Transvaal Museum (unpublished records); Entomology Department, South African Museum (unpublished records).

Doubtful localities, discussed in the literature, and single, highly disjunct records that could not be verified, were omitted. After screening for errors, the final species presence-only dataset (*sensu* Mugo *et al.* 1995) was submitted to the GIS for mapping and spatial analysis. A total of 6 766 data points were collated.

# 2.4 Spatial analysis

Digital distribution maps were produced for each species by superimposing point locality records on a dataset representing the political boundaries of southern Africa (south of 15° latitude), using ArcView GIS Version 3.2 (Environmental Systems Research Institute, Redlands, CA). A

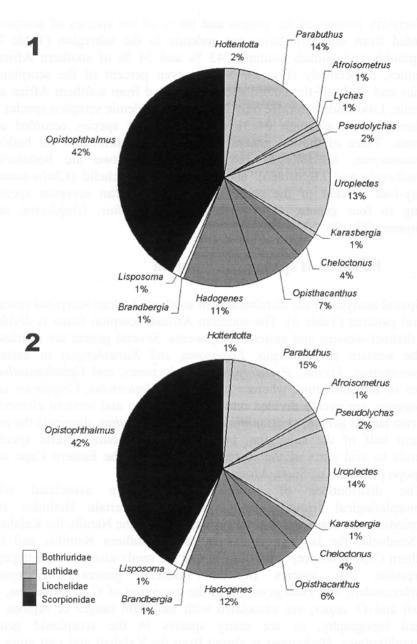
spatial join was then conducted by superimposing scorpion distributions on a dataset representing the QDS grid of southern Africa, to determine scorpion hotspots, i.e. areas of high species richness and endemism (Myers et al., 2000), at the scale of a QDS (Lombard, 1995a, 1995b). Hotspots were based on measures of species richness (all 140 species) and endemism (including only southern African endemics), which may reflect centres of endemism, or regions of speciation, given that most scorpion species are sedentary (Polis, 1990; Harington, 1984), and probably experience minimal range-shifting.

Spatial joins were also conducted to determine to what extent scorpion distributions are explained by topography, major sand systems and river drainage in southern Africa. A topographic contour dataset was created from the GTOPO30 raster grid, obtained from the website of the U.S. Government Public Information Exchange Resource: http://edcdaac.usgs. gov/gtopo30/gtopo30.html. The dataset of sand systems was created by clipping and merging relevant polygons extracted from a dataset of African geology from the Department of Marine Geoscience, University of Cape Town, with polygons extracted from a dataset of Namibian landforms from the Namibian National Biodiversity Task Force (Barnard, 1998), downloaded from: http://www.dea.met.gov.na/programmes/biodiversity/ countrystudy.htm. A da-taset of major rivers in southern Africa was created by merging polygons extracted from a dataset of Namibian rivers from the website of the Namibian National Biodiversity Task Force (Barnard, 1998) with a dataset of South African rivers from the Surface Water Resources of South Africa 1990 (WR90) database (Midgley et al., 1994), of the Water Research Commission, South Africa.

## 3. RESULTS

# 3.1 Composition of the fauna

Scorpionidae and Buthidae respectively comprise 42 % and 34 % of the southern African scorpion species, whereas Liochelidae and Bothriuridae respectively comprise 22 % and 2 % thereof (Fig. 1; Table 1). Despite a similar proportion of species, buthids comprise 54 % of the southern African scorpion genera, whereas scorpionids comprise only 8 %; liochelids and bothriurids respectively comprise 22 % and 15 % thereof. Eighty-one percent of the southern African scorpion species belong to four genera: Opistophthalmus, Parabuthus, Uroplectes, and Hadogenes.



*Figures 1,2.* 1. Proportion of species in the families and genera of southern African scorpions. 2. Proportion of endemic species in the families and genera of southern African scorpions.

Forty-six percent of the genera and 96 % of the species of scorpions recorded from southern Africa are endemic to the subregion (Table 2). Scorpionids and buthids comprise 43 % and 34 % of southern African endemics, respectively (Fig. 2). Ninety-seven percent of the scorpionid species and 96 % of the buthid species recorded from southern Africa are endemic. Liochelids comprise only 21 % of the endemic scorpion species in southern Africa, although 94 % of the liochelid species recorded are endemic. Three of the six genera endemic to the subregion are buthids (Afroisometrus, Karasbergia and Pseudolychas), two are bothriurids (Brandbergia and Lisposoma), and the last is a liochelid (Cheloctonus). Eighty-four percent of the endemic southern African scorpion species belong to four genera: Opistophthalmus, Parabuthus, Uroplectes, and Hadogenes (Fig. 2).

## 3.2 Patterns of species distribution

Spatial analysis of the distributions of southern African scorpions reveals several patterns (Table 3). The southern African scorpion fauna is divided into distinct western and eastern components. Several genera are restricted to the western (*Brandbergia*, *Lisposoma* and *Karasbergia*) or eastern (*Afroisometrus*, *Lychas*, *Pseudolychas*, *Cheloctonus*, and *Opisthacanthus*) halves of the subregion, whereas others, e.g. *Hottentotta*, *Uroplectes* and *Hadogenes*, are evenly divided into distinct eastern and western elements. The two largest genera, *Opistophthalmus* and *Parabuthus*, dominate the arid western half of the subregion, but each also contains several species endemic to arid parts of the eastern half, such as the Eastern Cape and Limpopo provinces of South Africa.

distributions of particular species are associated geomorphological features of the landscape. Certain Buthidae and Scorpionidae are restricted to sand systems such as the Namib, the Kalahari, the Sandveld, the isolated sand systems of southern Namibia and the Northern Cape Province, South Africa, and the sandy areas of the Limpopo Depression. species of the liochelid All genera Cheloctonus, Opisthacanthus, and Hadogenes, with the exception of C. crassimanus, C. jonesii and O. asper, are associated with mountain ranges or regions of rugged topography, as are many species in the scorpionid genus Opistophthalmus. Hadogenes is absent from the Kalahari and vast areas of the Cape Middleveld, the Free State, and the Karoo, as well as from the Springbok flats of the Limpopo Province, the sandy Mozambique plains and Makatini flats of northern KwaZulu-Natal Province, and the sandy areas of

Percentages of the total are provided Table 1. Scornion diversity in southern African countries

| Family       | Genus                        | Sth. Africa | V          | В          | T        | Ma      | Mo      | z       | SA       | S      | Za      | Zi      | Э       |
|--------------|------------------------------|-------------|------------|------------|----------|---------|---------|---------|----------|--------|---------|---------|---------|
| Bothriuridae | Brandbergia                  | 1(1)        | 13         | 0          | li<br>Ză | adi     |         | 1 (100) | 1        |        |         |         |         |
|              | Lisposoma                    | 2(1)        |            |            |          |         |         | 2 (100) |          |        |         |         |         |
|              | Total                        | 3 (2)       |            |            |          |         |         | 3 (100) |          |        |         |         |         |
| Buthidae     | Afroisometrus                | 1(1)        | E<br>Silon |            |          |         |         | 3       |          |        |         | 1 (100) |         |
|              | Hottentotta                  | 3 (2)       | 1 (33)     | 1 (33)     |          | 3       | 1 (33)  | 2 (67)  |          |        | 1 (33)  | 1 (33)  | 1 (33)  |
|              | Karasbergia                  | 1(1)        |            |            |          |         |         | 1 (100) |          |        | ,       |         | oin     |
|              | Lychas                       | 1(1)        |            |            |          | 1 (100) |         | 10      |          |        | 1 (100) | 1 (100) | 1 (100) |
|              | Parabuthus                   | 20 (14)     | 4 (20)     | 7 (35)     |          |         |         | 15 (75) |          |        | 3 (15)  | 5 (25)  |         |
|              | Pseudolychas                 | 3 (2)       |            | lon<br>orl |          |         | 1 (33)  |         |          | 1 (33) |         |         |         |
|              | Uroplectes                   | 19 (14)     | 3 (16)     |            | 2 (11)   |         | 6 (32)  | 10 (53) |          | 4 (21) | 4 (21)  | 6 (32)  |         |
|              | Total                        | 48 (34)     | 8 (17)     | 13 (27)    | 2(4)     | 2(4)    | 11 (23) | 28 (58) |          | 5 (10) | 9 (19)  | 14 (29) | 2 (4)   |
| Liochelidae  | Cheloctonus                  | 5 (4)       |            |            | 9        |         | 1 (20)  |         | 5 (100)  | 1 (20) | 5       |         |         |
|              | Hadogenes                    | 16 (11)     | 1 (6)      | 2 (13)     |          | 6       | 2(13)   | 4 (25)  |          | 1 (6)  | 1 (6)   | 2 (13)  |         |
|              | Opisthacanthus               | 10 (7)      |            | 1 (10)     | 2 (20)   |         | 2 (20)  |         |          | 3 (30) | 6       | 2 (20)  | 2 (20)  |
|              | Total                        | 31 (22)     | 1(3)       | 3 (10)     | 2 (6)    | 2 (6)   | 5 (16)  | 4 (13)  | 23 (74)  | 5 (16) | 1(3)    | 4 (13)  | 2 (6)   |
| Scorpionidae | Scorpionidae Opistophthalmus | 59 (42)     | 5 (8)      | (01) 9     | 1 (2)    |         | 3 (5)   | 28 (47) | 39 (66)  | 3      | 2(3)    | 4(7)    | 2(3)    |
| Total        | Genera                       | 13          | 5 (38)     | (94)       | 3 (23)   |         | (69) 6  | 8 (62)  | 11 (85)  | 5 (38) | (46)    | 7 (54)  | 7 (54)  |
|              | Species                      | 140         | (9) 6      | 22 (16)    | 5 (4)    |         | 19 (14) | 63 (45) | 100 (71) | 10 (7) | 12 (9)  | 22(16)  | (4)     |

*Table 2.* Scorpion endemism in southern African countries, south of 15° latitude. Percentages of the total are provided in parentheses. Abbreviations as follows: A (Angola); N (Namibia); SA (South Africa).

| Family       | Genus           | Sth. Africa | A    | N       | SA      |
|--------------|-----------------|-------------|------|---------|---------|
| Bothriuridae | Brandbergia     | 1(1)        |      | 1 (100) |         |
|              | Lisposoma       | 2(1)        |      | 2 (100) |         |
|              | Total           | 3 (2)       |      | 3 (100) |         |
| Buthidae     | Afroisometrus   | 1(1)        |      |         |         |
|              | Hottentotta     | 2(1)        |      |         |         |
|              | Karasbergia     | 1(1)        |      |         |         |
|              | Parabuthus      | 20 (15)     |      | 5 (25)  | 3 (15)  |
|              | Pseudolychas    | 3 (2)       |      | `       | 2 (67)  |
|              | Uroplectes      | 19 (14)     |      | 3 (16)  | 4(21)   |
|              | Total           | 46 (34)     |      | 8 (17)  | 9 (20)  |
| Liochelidae  | Cheloctonus     | 5 (4)       |      | 3 9     | 4 (8)   |
|              | Hadogenes       | 16 (12)     |      | 1 (6)   | 8 (5)   |
|              | Opisthacanthus  | 8 (6)       |      |         | 3 (3)   |
|              | Total           | 29 (22)     |      | 1 (3)   | 15 (52) |
| Scorpionidae | Opistophthalmus | 57 (43)     | 1(2) | 19 (32) | 27 (46) |
| Total        | Genera          | 6 (46)      |      | 2 (15)  |         |
|              | Species         | 134 (96)    | 1(1) | 31 (22) | 51 (36) |

the Namib. The scorpion faunas occurring to the north and south of the Orange River are also distinctly different, sharing only 20 species that occur on both sides

Most southern African scorpion species show discrete restricted distributional ranges (less than 50 QDS), with the exception of a few widespread species, e.g. P. granulatus, O. carinatus and O. wahlbergii. The distributional ranges of closely related species are invariably allopatric or parapatric, especially in the non-buthid genera Cheloctonus, Opisthacanthus, Hadogenes, and Opistophthalmus. The distributions of and Hadogenes are Opisthacanthus almost mutually exclusive, Opisthacanthus occupying the length of the eastern escarpment and the Cape Fold Mountains (excluding the Cedarberg) and Hadogenes occupying the interior plateau and the Cedarberg.

## 3.3 Hotspots

Hotspot analysis of species richness indicates that most parts of southern Africa contain at least one scorpion species (Fig. 3). The apparent absence of scorpions from large parts of Mozambique, northern Zimbabwe, and the central Kalahari in Botswana is a sampling artefact. Despite the bias caused by undersampling in these areas, coverage of southern Africa is fairly complete for an arthropod group.

Africa; north, south or extending across the Orange River; associated with major sand systems or mountain ranges; and intersecting up to 400 quarter-degree Afroisometrus (A); Lychas (Ly); Pseudolychas (Ps); Hottentotta (Ho); Karasbergia (K); Parabuthus (Pa); Uroplectes (U); Cheloctonus (C); Opisthacanthus Table 3. Patterns of scorpion distribution in southern Africa, expressed as the number of species with distributions in the west, east or centre of southern squares (QDS), an index of range size. Percentages of the total are provided in parentheses. Abbreviations as follows: Brandbergia (B); Lisposoma (Li); (Oh); Hadogenes (Ha); Opistophthalmus (Oo).

|              |                  | B, Li | A, Ly, Ps     | Ho      | Х    | Pa      | Ω     | C, Oh           | Ha     | 00      | Total   |
|--------------|------------------|-------|---------------|---------|------|---------|-------|-----------------|--------|---------|---------|
| Sth. Africa  | Western          | 3(2)  | E<br>IA<br>OV | 2(1)    | 1(1) | 15 (11) | 8 (6) | 0)<br>10)<br>18 | 6 (4)  | 43 (31) | 78 (56) |
|              | Eastern          |       | 5 (4)         | 1(1)    |      | 3 (2)   | (9) 6 | 15(11)          | (9) 6  | 10 (7)  | 52 (37) |
|              | Central          |       |               | ed<br>a |      | 3 (2)   | 2(1)  | V V             | q      | 6 (4)   | 11 (8)  |
| Sand Systems | Namib            |       |               |         |      | 4 (3)   |       |                 |        | 7(5)    | 11 (8)  |
| i l          | Kalahari         |       |               |         |      | 5 (4)   |       |                 |        | 4(3)    | (9) 6   |
|              | S Namibia-N Cape |       |               | 1(1)    |      | 4(3)    |       |                 |        | 4(3)    | (9) 6   |
|              | Sandveld         |       |               |         |      |         |       |                 |        | 4(3)    | 4 (3)   |
|              | Limpopo          |       |               |         |      | 3 (2)   |       |                 |        | 2(1)    | 5 (4)   |
| Mountains    | Escarpment       |       | 1(1)          |         |      | 2(1)    |       | 7 (5)           | (9) 6  | (9) 6   | 28 (20) |
|              | Cape Fold        |       |               |         |      |         | 3(2)  | 4(3)            | 1(3)   | 7(5)    | 15(11)  |
|              | Other            | 2(1)  |               | 1(1)    |      | 2(1)    | 1(1)  | 1(1)            | (9) 8  | 12(9)   | 27 (19) |
| Drange River | North            | 3(2)  |               | 1(1)    |      | (9) 8   | 5 (4) | OC<br>Ni<br>in  | 2(1)   | 24 (17) | 43 (31) |
|              | South            |       |               |         |      | 3 (2)   | 4(3)  |                 | 3(2)   | 22 (16) | 32 (23) |
|              | Transriverine    |       |               | 1(1)    | 1(1) | (9) 8   | 2(1)  |                 | 2(1)   | 6(4)    | 20 (14) |
| Range size   | 1-20             | 2(1)  | 2(1)          | 1(1)    |      | 7(5)    | 7 (5) | 10 (7)          | (9) 6  | 36 (26) | 74 (53) |
|              | 20-50            | 1(1)  | 3(2)          |         | 1(1) | 2(1)    | 3(2)  | 2(1)            | 2(1)   | 16(11)  | 30 (21) |
|              | 50-100           |       |               | 2(1)    |      | 7(5)    | 5 (4) | 3(2)            | 3(2)   | 4(3)    | 24 (17) |
|              | 100-200          |       |               |         |      | 4(3)    | 2(1)  |                 | 100    |         | 7(5)    |
|              | 200-300          |       |               |         |      |         | 2(1)  |                 | a<br>O | 2(1)    | 4(3)    |
|              | 300-400          |       |               |         |      | 1(1)    |       |                 |        | 3       | 2(1)    |

Twenty-four primary hotspots (11-16 species) occur in the western third of southern Africa. Most are concentrated in northwestern, central, and southern Namibia, and the Richtersveld and Namaqualand regions of the Northern Cape Province, South Africa, with an isolated hotspot in the Breede River Valley, Western Cape Province, South Africa. Secondary hotspots (7-10 species) are concentrated in the same parts of Namibia, as well as the Richtersveld, Namaqualand, and the Western Cape. Additional secondary hotspots occur in Bushmanland and the southern Kalahari, Northern Cape Province, South Africa, the Eastern Cape, Limpopo and Mpumalanga provinces of South Africa, and the Eastern Highlands of Zimbabwe.

Hotspots of endemic species show a similar pattern (Fig. 4). Six primary hotspots (9-12 endemics) occur along the Aus Mountains and the Huib-Hoch Plateau of southern Namibia, the western escarpment, Namaqualand and Breede River Valley in the Northern and Western Cape provinces of South Africa, and the Eastern Cape Province. Secondary hotspots (6-8 endemics) are concentrated in central and southern Namibia, the Richtersveld, Namaqualand and Bushmanland, the Western Cape, Eastern Cape, Limpopo and Mpumalanga provinces.

Most regions of southern Africa contain at least one endemic species, except for Mozambique, northern Zimbabwe, and most of the Kalahari (no scorpion species are endemic to Botswana). The absence of endemics from northern Zimbabwe and the Kalahari cannot be attributed simply to collector bias, however, as some of these areas, particularly eastern Namibia, northern Botswana, and the northeastern border of Zimbabwe have been fairly well sampled. The distributions of several species in these areas extend beyond the boundaries of southern Africa.

Two trends will be noted from the hotspot analyses. Firstly, most hotspots occur in the arid western half of southern Africa, or in arid parts of the eastern half, e.g. the Eastern Cape and the Lowveld of the Limpopo and Mpumalanga provinces, suggesting the influence of climate. Secondly, all hotspots occur in regions of rugged topography, complex geology, or substratal heterogeneity. The Namibian hotspots occur along the Skeleton Coast, the mountainous Kaokoveld and Damaraland, at the interface of the Central Namib Sand Sea and the western escarpment, and in the highlands on the interior plateau (e.g. the Khomas Hochland, the Karasberg, and the Huib-Hoch Plateau). The South African hotspots in the mountainous Richtersveld, Namaqualand and Bushmanland, the sand dunes of the southern Kalahari, and the western escarpment are further evidence of this pattern. Remaining hotspots are associated with the Cape Fold Mountains, the escarpment in Mpumalanga and the Eastern Highlands of Zimbabwe, the Lebombo Mountains, the Magaliesberg, the Waterberg-Soutpansberg

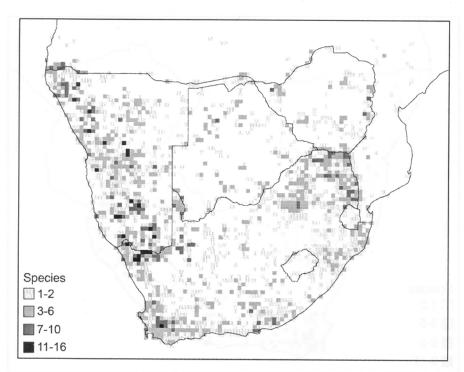


Figure 3. Hotspots of scorpion species richness in Africa, south of 15° latitude, expressed as the total number of species per quarter-degree square.

complex, and the sandy areas of the Limpopo Depression. The hotspot representing the Breede River Valley occurs at the junction of the north-south and east-west axes of the Cape Fold Mountains, and at a transition zone between fynbos and karoo vegetation types, respectively associated with higher and lower rainfall regimes. Notwithstanding the bias caused by undersampling in central Botswana and Mozambique, hotspots are in general poorly represented in areas of uniform topography, geology, and substratum, e.g. the Karoo, the Highveld, the central and northern Kalahari, and the Mozambique plain.

#### 4. DISCUSSION

## 4.1 Models of speciation

Biogeography should explain the distribution of extant taxa in terms of historical factors together with the use of their contemporary ecology 40 Lorenzo Prendini

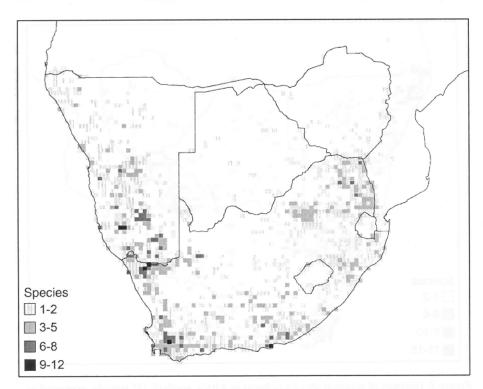


Figure 4. Hotspots of scorpion endemism in Africa, south of 15° latitude, expressed as the number of endemic species per quarter-degree square.

(Myers and Giller, 1988; Haydon et al., 1994). As fossil evidence is absent, biogeographic considerations of southern African scorpions must be derived primarily from ecological sources (Lamoral, 1978b). This approach is considered valid, however, as the habits of scorpions appear to have altered little over time (Jeram, 1990b; Sissom, 1990). The high species richness and endemism of southern African scorpions may be attributed to two main factors: a variable climate and a topographically diverse landscape (associated with complex geology). These factors, together with the specialised ecological requirements of most scorpion species, appear to have promoted speciation, especially during periods of palaeoclimatic change.

Speciation is most often explained by the allopatric model: species evolve from components of larger populations isolated in refugia by a cessation of gene flow between them (Mayr, 1942, 1963). Cessation of gene flow between populations may result from dispersal or vicariance. Although the existence of a physical barrier to gene flow is implied by both, vicariance and dispersal constitute distinct models of disjunction differentiated as follows: vicariance implies that the splitting of an ancestral population was caused by the appearance of a barrier; dispersal implies that

the splitting occurred by movement across a pre-existing barrier (Platnick and Nelson, 1978). Neither dispersal nor vicariance explanations should be discarded *a priori* as irrelevant for any particular group of organisms. Biogeographic analysis should allow us to choose objectively between these two types of explanations for particular taxa.

Scorpion dispersal is limited to terrestrial vagility. As aquatic and aerial dispersal are improbable, barring unusual circumstances such as transoceanic rafting and synanthropy (Newlands, 1973, 1978a), the biogeography of scorpion faunas separated by barriers is best explained through a vicariance model, terrestrial dispersal in turn being controlled at any time by various ecological requisites (Lamoral, 1978b). Given the specialised ecological requirements of southern African scorpions, it is feasible to construct a scenario of how repeated speciation might have produced current levels of diversity and endemism in the subregion, in a similar manner to that proposed by Koch (1977, 1981) for the scorpions of Australasia. The remainder of this paper is devoted to a discussion of the ecological and historical factors determining the distribution of southern African scorpions, insofar as these account for their diversity and endemism.

## 4.2 Ecological factors

#### 4.2.1 Climate

Different genera and species of scorpions have adapted to xeric and mesic habitats in southern Africa, as elsewhere. At the broadest level, the distinct western and eastern elements of the southern African fauna confirm the importance of climate as a determinant of scorpion distribution in the subregion. Two components of climate are thought to affect scorpion distribution, viz. rainfall (relative humidity) and temperature. Rainfall has a noticeable effect on scorpion distribution in Israel (Warburg and Ben-Horin, 1978; Warburg et al., 1980), whereas temperature appears to be the primary factor limiting the southward expansion of tropical scorpion species on the east coast of Australia (Koch, 1977, 1981). The absence of scorpions from higher latitudes may be indicative of the importance of temperature on a global scale (Gromov, 2001); few species occur where snow is a common occurrence (Williams, 1987; Polis, 1990).

Previous authors (Hewitt, 1925; Lawrence, 1942, 1952; Newlands, 1978a, 1978b) observed that several scorpion genera are restricted to, or dominant in, the arid to semi-arid western half of southern Africa, whereas others are restricted to the humid east, a pattern confirmed in the present investigation. The distributions of *Hottentotta* and *Parabuthus* have been related to the 600 mm isohyet, as these genera are generally absent from

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areas receiving more than 600 mm of annual rainfall (Newlands, 1978b). In contrast, Afroisometrus, Lychas, Pseudolychas, Cheloctonus and Opisthacanthus require high rainfall, and several species in these genera are endemic to Afromontane forests (Lawrence, 1942, 1953). Their limited distributions may be partly indicative of their relative humidity requirements. Lisposoma joseehermanorum is restricted to humid subterranean habitats such as caves, and leaf litter under boulders on south and east-facing slopes in the Otavi Highlands, which receive the highest rainfall in Namibia (Lamoral, 1978b, 1979; Prendini, 2003b). The restricted distributions of certain silvicolous Uroplectes (e.g. U. insignis on the forested eastern slopes of Table Mountain) may likewise be related to their narrow climatic requirements.

The distinction between the western and eastern components of the southern African scorpion fauna is repeated within more widespread genera such as *Opistophthalmus*, *Parabuthus*, *Hottentotta*, *Hadogenes* and *Uroplectes*, each of which contains particular species or groups of related species that are restricted to either the humid east or the arid west of southern Africa. Newlands (1978a: 688) stated "water is probably the most important single factor affecting the distribution of scorpions in southern Africa in that it is largely responsible for the type of vegetation in any given region and, to an extent, the daily temperature fluctuation of the soil surface."

#### 4.2.2 Substratum

The importance of the substratum in scorpion ecology and distribution is well known (San Martín, 1961; Smith, 1966; Lamoral, 1978a, 1978b, 1979; Bradley, 1986, 1988; Bradley and Brody, 1984; Polis and McCormick, 1986; Fet et al., 1998, 2001; Prendini, 2001a). Lamoral (1978a, 1978b, 1979) established that the distribution of *Opistophthalmus* in southern Africa is determined primarily by soil hardness and, to a lesser degree, by soil texture, each species being restricted to soils within a certain range of hardness, rather than to a particular soil type. Lamoral (1978b: 305) concluded that "the nature of the substratum, taken in its broadest possible definition, is probably the most important single factor that has and still determines the distribution of scorpions ... the nature of the substratum is affected to a greater or lesser extent by vegetation, which is in turn partly the result of prevailing climatic conditions."

Newlands (1978a) and Lamoral (1979) independently classified the southern African scorpions according to their habitat predilections, both recognising a distinction between arboreal, rock-dwelling and burrowing species. Rock-dwelling species were further subdivided into species that inhabit crevices and species that shelter under stones, whereas burrowing

species were subdivided according to their occurrence in soft or hard substrata. Bradley (1988) and Polis (1990) redefined these ecomorphotypes for all scorpions (see also Tikader and Bastawade, 1983; Fet et al., 1998). Prendini (2001a) introduced the concept of substratum-specialization and identified stenotopic vs. eurytopic ecomorphotypes, which were then applied to the southern African scorpion species.

Most species of Parabuthus, Opistophthalmus, and Cheloctonus are fossorial. They are divided into psammophilous and pelophilous species (Lawrence, 1969; Newlands, 1972a; Prendini, 2001a). Psammophilous and semi-psammophilous species of Parabuthus and Opistophthalmus display ecomorphological adaptations to increase locomotor and burrowing efficiency in loose sand (Lawrence, 1969; Newlands, 1972a, 1978a; Polis, 1990; Prendini, 2001a). These adaptations are exaggerated in ultrapsammophilous species (Newlands, 1972a) that occupy shifting sand dune environments (e.g. O. flavescens and O. holmi). All psammophilous scorpions are specialists, poorly adapted to life outside their sandy habitats. They are unable to burrow in harder or coarser substrata (Polis, 1990), and their distributions are restricted to sand systems. In southern Africa, such habitats occur in the northern and southern Namib, the Kalahari, southern Namibia, the Northern Cape Province, and the Limpopo Province. As species that burrow in soft, sandy soils tend to be restricted to a smaller range of substratum hardness than those burrowing in harder substrata, only a few (e.g. P. granulatus) are widely distributed, tracking generalised sandy environments across southern Africa.

Pelophilous species burrow in sandy loam and clay soils, and display ecomorphological adaptations to assist with loosening these compacted substrata (Newlands, 1972a, 1972b, 1978a; Polis, 1990; Prendini, 2001a). Pelophilous species of *Opistophthalmus* are cheliceral burrowers (Newlands, 1972b; Eastwood, 1978b). In contrast, *Cheloctonus jonesii*, a liochelid common in black turf soils (Newlands, 1978a), is a pedipalpal burrower that uses its large, rounded pedipalp chelae for burrowing in this extremely hard clayey substratum (Harington, 1978). Several pelophilous species, e.g. *O. glabrifrons*, are widely distributed, presumably because they are able to burrow in a greater range of soil hardness than is possible for psammophilous species.

Substratum hardness is carried to the extreme in the rock habitats of lithophilous scorpion species, specialists adapted to life in the narrow cracks and crevices of rocks (Newlands, 1972b, 1978a; Polis, 1990; Prendini 2001a, 2001b). The ecomorphological adaptations that characterise lithophilous scorpions facilitate locomotion on rock but hinder locomotion on other substrata. These scorpions are therefore usually restricted to regions of mountainous or rugged topography, their distributions often closely matching the boundaries of mountain ranges and geological

formations (Newlands, 1972b, 1978a, 1980; Prendini, 2001b). Lithophilous adaptations are exaggerated in the southern African genus Hadogenes, all species of which inhabit the cracks and crevices of weathered outcrops consisting usually of fine-grained quartzite or igneous rocks such as granite, norite, syenite, diorite, and gabbro (Newlands, 1980). Hadogenes are absent from vast areas of southern Africa that are either rockless or contain unsuitable geology. Most Cheloctonus and Opisthacanthus are generalist lithophiles, sheltering under rocks or logs, in addition to occupying crevices (Lawrence, 1953; Newlands, 1972b). Their requirement for higher humidity appears to have restricted their distributions to the eastern escarpment, the Cape Fold Mountains, and the southern and eastern coastal plains. Hadogenes, which tolerate lower rainfall, are more widely distributed in the interior (Lawrence, 1942; Newlands, 1980). Males of certain Opistophthalmus that excavate shallow scrapes under rocks, or between slabs of rock (e.g. O. austerus, O. karrooensis, and O. pallipes), show similar ecomorphological adaptations to *Hadogenes* (Eastwood, 1978b; Prendini, 2001a). Such semi-lithophilous species are characterised by discrete, allopatric distributions in particular mountain ranges.

Only four obligate corticolous species, that shelter in holes or under the loose bark of trees, often several metres above the ground, occur in southern Africa (Lamoral, 1979; Newlands, 1978a; Newlands and Martindale, 1980; Prendini, 2001b): Lychas burdoi; Uroplectes otjimbinguensis; U. vittatus; Opisthacanthus asper. Corticolous species display few ecomorphological adaptations and are widely distributed. All except U. otjimbinguensis are restricted to the eastern half of southern Africa.

Lapidicolous scorpions, which shelter under stones or any other available cover, are habitat generalists (Newlands, 1978a; Lamoral, 1978b, 1979; Prendini, 2001a), displaying few ecomorphological adaptations and varied, often widespread distributions (e.g. *U. planimanus* and *U. triangulifer*), governed primarily by climate. In southern Africa, most species of *Hottentotta*, *Pseudolychas*, and *Uroplectes* are lapidicolous. Several normally ground-dwelling species occasionally shelter in arboreal habitats (e.g. *H. conspersus*, *P. villosus*, *U. formosus*, *U. insignis*, *U. lineatus*, and *U. olivaceus*) or are epigeic on vegetation while foraging nocturnally (e.g. *P. villosus* and *U. gracilior*) (Lamoral, 1978b, 1979).

#### 4.2.3 Biotic interactions

Scorpion-scorpion interactions affect the distributions of many scorpion species (Williams, 1970, 1980; Koch, 1977, 1978, 1981; Lamoral, 1978a, 1978b; Shorthouse and Marples, 1980; Bradley and Brody, 1984; Polis and McCormick, 1986, 1987). The exact nature of these interactions is controversial, two main hypotheses with similar predictions having been

presented (Polis, 1990): exploitation competition for prey or homesites and interference, manifested as aggression or intraguild predation among potential competitors. Exploitation competition for limited resources selects for ecological divergence of species from one another in resource use, whereas interference competition selects for subordinate species' avoidance of dominant species (Simberloff, 1983). Divergence and avoidance may decrease habitat overlap and even produce competitive exclusion in certain regions, thereby resulting in geographical allopatry or parapatry; e.g. in Australia, *Urodacus* Peters, 1861 species are almost totally absent where Liocheles waigiensis (Gervais, 1843) is abundant (Koch, 1977, 1981). Alternatively, the decrease in habitat overlap caused by divergence or avoidance may allow coexistence, resulting in sympatry; e.g. *Hadrurus* arizonensis Ewing, 1928, Paruroctonus luteolus (Gertsch and Soleglad, 1966), Paruroctonus mesaensis (Stahnke, 1957), and Vaejovis confusus Stahnke, 1940 in California (Polis and McCormick, 1986, 1987). In addition, intraguild predation may allow coexistence by preventing exclusion of competitively inferior species through differential exploitation of competitively superior species (Polis and Holt, 1992).

Competition theory predicts that coexistence, and thus sympatry, will be associated with niche differences and "resource partitioning" (Simberloff, 1983): decreased overlap in the use of critical resources, different spatiotemporal patterns, and differences in body size. Evidence suggests that niche differences exist in space and time, but not in the use of food (Polis, 1990). Although there are exceptions (Main, 1956; Koch, 1977, 1981; Lourenço, 1983b), most scorpion species are generalists, eating any prey they are able to capture (Polis, 1979; McCormick and Polis, 1990). Size differences may divide prey eaten by adults of different sizes. For example, in southern Africa, sympatric species of Opistophthalmus often differ in size (e.g. O. flavescens and O. holmi of the Namib, or O. wahlbergi and O. concinnus of the Kalahari), as do sympatric species of *Hadogenes* (e.g. *H. tityrus* and *H.* taeniurus or H. zumpti), implying a difference in ecological niche. However, as adults of large species must grow from a small size at birth, all species overlap in size (and associated size-related prey use) during some part of their lives and such developmental overlap greatly limits the effectiveness of body size differences to divide resources among those species that show a wide range in size during development (Polis, 1990).

Size is nonetheless of paramount importance in interference among scorpions. Intraguild predation is known to occur among at least 30 pairs of scorpion species at six North American sites (Polis et al., 1981; Bradley and Brody, 1984; Polis and McCormick, 1986, 1987; Polis, 1990), and has personally been recorded in southern Africa between *Parabuthus granulatus* and *Opistophthalmus wahlbergi*, *Parabuthus transvaalicus* and *Hadogenes troglodytes*, *H. troglodytes* and *Opistophthalmus boehmi*, *H. troglodytes* and

Opistophthalmus glabrifrons. In California, the impact of intraguild predation by Paruroctonus mesaensis and Hadrurus arizonensis is reflected in niche shifts among the smaller prey species (Paruroctonus luteolus and Vaejovis confusus), which are relatively uncommon in the habitats occupied by the intraguild predators (Polis and McCormick, 1986, 1987). The subordinate species occupy microhabitat refuges, avoiding ground predation by foraging primarily from burrow entrances (P. luteolus) or on vegetation (P. luteolus and V. confusus). This is also seen in southern Africa, where smaller species of Uroplectes and Opistophthalmus, as well as juveniles of larger species (e.g. Parabuthus) forage on vegetation, avoiding predation by larger Parabuthus and Opistophthalmus, including adult conspecifics.

The effect of substratum hardness is an important spatial factor governing the coexistence of different scorpion species. Koch (1977, 1978, 1981) argued that the distribution of species of the fossorial Australian genus Urodacus is determined by competition for homesites, citing differential burrow construction among different species as evidence of decreased habitat overlap. Lamoral (1978a, 1978b) argued similarly that specific soil-hardness preferences decrease competition for burrow sites and allow coexistence among southern African Opistophthalmus. Species with overlapping hardness preferences tend to be allopatric or parapatric, sympatry usually occurring only when two or more species occupy substrata of very different hardness within the same geographical locality (in which case they are allotopic), as shown in the Kalahari species O. carinatus and O. wahlbergi, which burrow in calcrete and aeolian sand, respectively (Lamoral, 1978a, 1978b). This may, in part, account for the discrete distributions of most species of *Opistophthalmus*. Sympatric species of *Opistophthalmus* may also coexist by differential burrow location: in northern Namaqualand, O. granifrons burrows on sandy-loam flats at the base of inselbergs where *O. peringueyi* occupies a semi-lithophilous niche under stones. Similarly, *O. boehmi* burrows in sandy-loam flats at the base of the northern slopes of the Soutpansberg, where O. lawrencei burrows under stones on the lower slopes (Newlands, 1969).

More fossorial buthid species are observed in sympatry than is the case among non-buthids, but this may be due to more subtle size differences and spatio-temporal microhabitat utilisation. In the southern Kalahari, five fossorial *Parabuthus* species (*P. granulatus*, *P. kalaharicus*, *P. kuanyamarum*, *P. laevifrons*, and *P. raudus*) coexist on the same sand dunes by means of differences in size and spatial occurrence on the dune. Sympatric scorpion species may also differ in foraging station: some hunt in vegetation, others forage on the ground, some "doorkeep" at their burrow entrance; some move constantly during foraging, and others are sit-and-wait foragers (Polis, 1990). In the Kalahari, *P. granulatus*, *P. kalaharicus* and *P.* 

kuanyamarum are active ground-surface foragers, P. laevifrons is epigeic on vegetation, and P. raudus is a sit-and-wait forager.

Presumably, exploitation competition for the restricted, two-dimensional rock crevice habitat, or interference through intraguild predation by the larger Hadogenes has prevented coexistence with the lithophilous species of Cheloctonus and Opisthacanthus, the distributions of which are almost entirely allopatric with Hadogenes. Exceptions are observed only when there are niche differences. For example, the pelophilous C. jonesii is sympatric with the lithophilous H. newlandsi and the corticolous O. asper in Limpopo Province, South Africa. Generally, species of *Hadogenes* occupy hotter, drier habitats than the smaller lithophilous Opisthacanthus and Cheloctonus, suggesting that there is also a difference in microclimatic requirements between them. In the Mpumalanga Province, South Africa, H. bicolor and O. validus occur in sympatry (e.g. at Blyderivierspoort and Haenertsburg), but they are allotopic, H. bicolor inhabiting exposed rock outcrops and O. validus inhabiting outcrops under forest cover (Newlands, 1980). However, climatic differences fail to explain the replacement, by Hadogenes minor in the Cedarberg, of Opisthacanthus, which occurs throughout the rest of the Cape Fold Mountains, where geology and climate are largely similar. Exploitation competition or intraguild predation seems a more plausible explanation. Almost all congeneric species of Hadogenes, Cheloctonus and Opisthacanthus occupy discrete, allopatric or parapatric distributional ranges, presumably for the same reason (Newlands, 1980). Exceptions are observed only when there are niche differences. For example, the pelophilous C. jonesii, lithophilous O. laevipes and corticolous O. asper are sympatric in Swaziland and the Mpumalanga and KwaZulu-Natal provinces, South Africa.

## 4.3 Historical factors

#### 4.3.1 Continental drift

As Gondwanaland fragmented, each of the southern continents carried with it a sample of the ancestral biota. This is well illustrated by several groups (MacArthur, 1972), including scorpions. Bothriurid scorpions, also known from South America, Australia and India, are represented in southern Africa by the genera *Brandbergia* and *Lisposoma* (Francke, 1982; Prendini, 2000a, 2003a, 2003b). Each contains species endemic to northern Namibia that display distributions typical of deserticolous palaeogenic elements (Stuckenberg, 1962). Lamoral (1978b, 1979) suggested that *Lisposoma* is a relict of a formerly tropical forest-dwelling ancestral element that survived the onset of aridification in Miocene times by resorting to a semi-endogean

existence, and offered the eucdaphic habitat of L. joseehermanorum as supporting evidence. The subsequent adoption of a lapidicolous habitat by L. elegans may have contributed to its present broader distribution and occurrence in habitats considerably drier than those inhabited by L. joseehermanorum (Prendini, 2003b).

A similar evolutionary scenario is proposed for the monotypic buthid genus *Karasbergia*, a fossorial endemic of rocky desert habitats in southern Namibia and the Northern Cape Province, South Africa (Lamoral, 1978b, 1979; Prendini, in press). This species, possibly related to *Charmus* Karsch, 1879 from India and Sri Lanka (Prendini, in press; E.S. Volschenk, pers. comm.), may also be considered a deserticolous palaeogenic element.

Taking possible vicariance and dispersal into account, few affinities exist between the scorpion faunas of the Afrotropical and Neotropical regions. There are, however, three notable exceptions. The first of these is the buthid genus *Ananteris* Thorell, 1891, with 19 species in Central and South America (including the island of Trinidad), and a single species from Guinea and Guinea-Bissau in West Africa, formerly placed in a monotypic genus, *Ananteroides* Borelli, 1911 (Gonzalez-Sponga, 1972, 1980; Lourenço, 1982, 1984b, 1985, 1987b, 1991c, 1993; Lourenço and Flórez, 1989; Fet and Lowe, 2000). The transfer of *Ananteroides feae* Borelli, 1911 to *Ananteris* by Lourenço (1985) was not justified phylogenetically and the monophyly of *Ananteris* remains to be tested.

The second exception is the diplocentrid genus Heteronebo Pocock, 1899, currently comprising 15 species. Thirteen species of Heteronebo are endemic to the Greater and Lesser Antilles in the Caribbean (Francke, 1978; Francke & Sissom, 1980; Armas, 1981, 1984, 2001; Stockwell, 1985; Armas & Marcano Fondeur, 1987; Prendini, 2000a; Sissom & Fet, 2000). However, two species inhabit Abd-el-Kuri Island off the Horn of Africa, east of Somalia and south of Yemen (Pocock, 1899; Francke, 1979; Sissom, 1990; Prendini, 2000a). Additional records of Heteronebo, probably representing undescribed species, were recently obtained from the nearby island of Socotra (W. Wranik & B. Striffler, pers. comm.). Whether the New and Old World species of Heteronebo are congeneric remains unclear. Prendini's (2000a) phylogenetic analysis confirmed the monophyly of Heteronebo based on the monophyly of a New and an Old World exemplar species of the genus. However, the grouping of New and Old World Heteronebo was weakly supported and may not withstand analysis with additional taxa and characters.

The liochelid genus *Opisthacanthus* (Lamoral, 1978b, 1980a) is the third exception. *Opisthacanthus* currently contains seven species in northern South America, southern Central America, the Cocos Islands off the coast of Costa Rica, and Hispaniola Island in the Caribbean, twelve species in Africa, and two species in Madagascar (Lourenço, 1980, 1981, 1987a, 1988,

1991a, 1995, 1997, 2001; Fet, 2000b; Lourenço & Fé, 2003). The origin of the Neotropical species is contentious. Newlands (1973, 1978a) proposed that they dispersed from Africa by trans-Atlantic rafting. However, a more parsimonious conclusion is that they are Gondwana relicts, which reached the Neotropics prior to the African disjunction in the late Cretaceous, and subsequently evolved in isolation (Francke 1974; Lamoral, 1980a; Lourenço, 1984a, 1989; Sissom, 1990). Both hypotheses rest on the assumption that Opisthacanthus is monophyletic, an assumption that was questioned on the basis of morphological evidence (Prendini, 2000a). Most of the African species of Opisthacanthus appear to be more closely related to other African liochelid genera (e.g., Cheloctonus), than to the Neotropical species. The African species of Opisthacanthus are not monophyletic either. Opisthacanthus lecomtei is more closely related to the South American species, placed in subgenus Opisthacanthus, whereas the others, traditionally placed in subgenus Nepabellus, are more closely related to Cheloctonus (Prendini, 2000a). The relationships of the Malagasy species of Opisthacanthus-for which a new subgenus, Monodopisthacanthus Lourenço, 2001, was recently erected—to the other species of the genus, also remain unclear. Determining the phylogenetic relationships of the species of Opisthacanthus to the other liochelid genera has implications for understanding the biogeography of Liochelidae as a whole.

Opisthacanthus and the related genus Cheloctonus conform closely to the criteria proposed for montane palaeogenic elements by Stuckenberg (1962), reiterating a distribution pattern displayed by many other groups of southern African invertebrates. Species of both genera occur in humid habitats at high altitude, on the south- and east-facing slopes of the Drakensberg escarpment (including the Eastern Highlands of Zimbabwe), and the Cape Fold Mountains. Lychas burdoi and the related buthids Afroisometrus and Pseudolychas have similar ecological requirements, and distributional ranges, suggesting that these may also be montane palaeogenic elements.

The genera *Brandbergia*, *Lisposoma*, *Afroisometrus*, *Karasbergia*, *Pseudolychas*, *Cheloctonus*, and *Opisthacanthus* are all relatively basal in their respective clades, and represent an ancient, relictual element in a fauna otherwise characterised by highly derived and speciose genera, e.g., *Parabuthus*, *Hadogenes*, and *Opistophthalmus*, which probably radiated fairly recently in post-Miocene or Pliocene times (Lamoral, 1978b, 1979; Prendini, 2000, 2001b, 2003a, 2003b, 2004b; Prendini et al., 2003).

## 4.3.2 Geomorphology and climate

Mountain ranges exert a significant influence on rainfall, temperature and vegetation, and may thus have indirectly promoted speciation in taxa

with restricted climatic tolerances during periods of palaeoclimatic change. For example, species of Cheloctonus, Opisthacanthus, Pseudolychas, and Uroplectes requiring high humidity probably experienced vicariance when formerly continuous patches of Afromontane forest or fynbos became isolated along the eastern escarpment during dry periods from the Pliocene to the Pleistocene (van Zinderen Bakker, 1962, 1976, 1978), and speciated in consequence. The sister species O. validus and O. lamorali, from the Drakensberg of South Africa and Eastern Highlands of Zimbabwe, respectively, separated by at least 300 km of dry savanna in the Limpopo Depression, are one of several examples. Speciation by vicariance associated with climate-induced expansion and contraction of montane forest or fynbos habitats (e.g. in "Pleistocene refugia") is the favoured explanation for high levels of endemism among montane palaeogenic elements (Stuckenberg, 1962). Most southern African species of Cheloctonus, Opisthacanthus, and Pseudolychas, and several southern African species of Uroplectes, may be products of such processes.

Expansion and contraction of the "arid corridor", connecting arid southwestern and northeastern Africa during successive wet and dry phases from the Pliocene to the Pleistocene (Balinsky, 1962; van Zinderen Bakker, 1969) could similarly have induced speciation among arid-adapted species, explaining the existence of *Parabuthus*, *Hadogenes* and *Opistophthalmus* in northeastern Africa (Prendini, 2001b, 2001c; Prendini et al., 2003).

Topography may also be responsible for speciation in scorpions by acting as barriers to dispersal (Koch, 1977). Some of the more important mountain barriers in southern Africa include the Great Escarpment, the Namaqua Highlands, Cape Fold Mountains, Lebombo Mountains, Magaliesberg, Waterberg, and Soutpansberg of South Africa, and the highlands of Namibia, e.g. the Khomas Hochland, Aus Mountains, Huib-Hoch Plateau, Otavi Highlands, and Karasberg. Many of these mountain ranges separate related scorpion species (Lamoral, 1978b).

There is also evidence that sand systems constitute barriers to non-psammophilous scorpions. Lamoral (1978b, 1979) hypothesised that, since the Pliocene, the Kalahari has acted as an agent of vicariance preventing migration of scorpion species along the "arid corridor". Lamoral (1978b) provided an example of how mountains and sand systems may have contributed to the present distribution of *Hottentotta*, a largely Palaearctic genus, in southern Africa. By the end of the Oligocene, the ancestral species of this genus had migrated as far south as their present distributions, facilitated by prevailing tropical and subtropical climates and vegetation. The emergence of the Kalahari sand system during the Pliocene (van Zinderen Bakker, 1975) induced the vicariance of the southwestern and northeastern species groups, resulting in the speciation of *H. trilineatus* in the northeast. Western expansion of the Kalahari forced the ancestral

southwestern group to migrate west. Once the western front of the Kalahari sands had reached the Central Highlands up to the 1 500 m contour, presumably in the Pleistocene when climate was generally colder, the ancestral range was effectively bisected and a vicariance established that caused the speciation of H. conspersus in the north and H. arenaceus in the south. A similar scenario was proposed to account for the distributions of various species of Parabuthus, Uroplectes, and Opistophthalmus north (P. gracilis, P. kraepelini, U. otjimbinguensis, U. planimanus, O. brevicauda, O. cavimanus, O. coetzeei, and O. ugabensis) and south (P. laevifrons, P. nanus, P. schlechteri, U. gracilior, U. longimanus, U. schlechteri, U. tumidimanus, O. fitzsimonsi, O. gigas, O. haackei, O. intercedens, O. opinatus and O. schultzei) of the Namibian Central Highlands (Lamoral, 1978b). This hypothesis rests on the assumption that the species in question are intolerant of sandy environments and fails to account for the distributions of semi-psammophiles such as P. gracilis, P. laevifrons, P. nanus, O. coetzeei, O. fitzsimonsi, and O. intercedens. Moreover, there is no reason to expect that, for the full duration of the Pleistocene, with its fluctuating warm and cold periods (van Zinderen Bakker, 1962, 1975, 1978), the Central Highlands would have constituted an ecological barrier to the ancestors of pelophilous Opistophthalmus which burrow under stones (e.g. O. cavimanus, O. opinatus and O. schultzei), or to species with semilithophilous adults (e.g. O. brevicauda, O. ugabensis, O. gigas, and O. haackei).

## 4.3.3 Substratum-specialization

Hotspots of species richness are located primarily in regions of complex topography and geology. The large number of species restricted to particular mountain ranges suggests that speciation often occurs in mountainous areas, rather than adjacent to them. Thus it appears as though intermontane valleys and depressions commonly act as barriers, rather than the other way around.

This may be related to the substratum-specialization of many scorpions, discussed extensively elsewhere (Prendini, 2001a). For example, the erosion of a mountain range, inhabited by a lithophilous scorpion species, would create isolated populations on the resultant inselbergs and ridges. Gene flow would cease once there were no longer rocky outcrops between adjacent inselbergs or ridges, owing to the substratum-specialization of the scorpion species. Observations for *Hadogenes* suggest that very narrow valleys or plains may interrupt gene flow between adjacent populations (Newlands, 1978a, 1980; Newlands and Cantrell, 1985). In the Pretoria area, *H. gunningi* inhabits the Magaliesberg and ridges to the south, whereas *H. gracilis* occurs on a series of ridges running parallel 2-3 km north; the narrow valley between them is free of rock outcrops, and thus acts as a

complete barrier to gene flow. The parapatric distribution patterns of *H. trichiurus* in the southern and eastern escarpment and *H. zuluanus* in the Lebombos reveal a similar process: the geomorphology of the hills in the area between differs in that the boulders are composed of basaltic rock, without cracks and crevices, and there are thus no suitable habitats for *Hadogenes* (Newlands, 1980).

Parts of mountain ranges may also have become separated by the invasion of aeolian sand during the Miocene or Pliocene (van Zinderen Bakker, 1975) leading to speciation in the isolated scorpion populations. For example, the Uri-Hauchab Mountains of the southern Namib are separated from the escarpment by a narrow belt of sand which blew in from the coast (Newlands, 1972c, 1978a, 1980). The isolation of these mountains probably took several million years, during which the isolated population of *Hadogenes* evolved into a new species, *H. lawrencei*, morphologically and genetically distinct from its sister species, *H. tityrus*, on the escarpment. As there are no rocks in the area between the Uri-Hauchab Mountains and the escarpment, the 26 km stretch of soft sand provides an insurmountable barrier to their dispersal.

Restricted ecological requirements probably account for the limited distributional ranges of most *Hadogenes*. Presumably, *Hadogenes*, which displays a suite of apomorphic character states, evolved from liochelid ancestors similar to *Opisthacanthus* and *Cheloctonus* (Lamoral, 1978b; Newlands, 1980), most of which occupy a similar, but more generalist lithophilous niche. Specialization in the lithophilous niche and the development of a greater tolerance for high temperatures and low rainfall may have enabled *Hadogenes* to exploit the arid to semi-arid interior of southern Africa, facilitating speciation by vicariance and, ultimately, producing a greater proportion of range-restricted endemics.

Speciation promoted by substratum-specialization may also be extended to fossorial taxa. Psammophilous and semi-psammophilous species, tracking deposits of a particular hardness, may have experienced vicariance when pockets of aeolian sand became separated from larger sand systems – a frequent occurrence during past periods of increased aridity such as the Pliocene, when wind action was believed to have transported vast quantities of sand over the western interior of southern Africa (van Zinderen Bakker, 1975; Lancaster, 1990). The occurrence of isolated populations of psammophilous species in areas of aeolian Kalahari sand support this notion: *P. granulatus*, *P. kuanyamarum* and *O. wahlbergii* occur in sandy areas northwest of the Soutpansberg, several hundred kilometers east of the main Kalahari sand system, but are not found on the intervening calcrete (Newlands, 1969, 1974).

Numerous similar examples of isolated psammophilous and semipsammophilous Parabuthus and Opistophthalmus populations occur in

Namibia, especially the sandy regions of the south, and in the Richtersveld and Namaqualand of South Africa, where numerous small dunes or dunefields, of mixed Namib or Kalahari sand origins, are isolated from the main sand systems against mountains or on sandless plains. Lamoral (1978b) suggested that psammophilous species probably evolved as a result of dispersal into the sand systems, rather than as a result of vicariance. Phylogenetic analyses show that psammophilous and semi-psammophilous species of Parabuthus, Uroplectes and Opistophthalmus are derived and must, therefore, have evolved after the sand systems became well established, speciating after dispersal into an environment that had previously constituted a barrier (Lamoral, 1978b, 1979; Prendini, 2001b). Post-Pliocene adaptation to the sandy substrata was proposed to account for species inhabiting the Kalahari, the Namib, and the sandy areas of southern Namibia and the Northern Cape Province, South Africa. Although it is certainly correct that the psammophilous and semi-psammophilous lineages in these genera evolved after colonising these sandy environments, the actual speciation events that gave rise to current species must have involved vicariance. If cessation of gene flow is an assumed prerequisite for allopatric speciation, the only manner in which populations of a psammophilous or semi-psammophilous scorpion species, unable to burrow in harder soils, could become isolated is through vicariance, i.e. by translocation on small sand dunes or dune-fields that gradually became separated from larger sand systems. Even in historical time, crescent dunes, e.g. barchans, have been found to travel considerable distances over sandless plains, carrying their associated arthropod fauna with them (Penrith, 1979; Endrödy-Younga, 1982). Speciation by vicariance among psammophilous and semipsammophilous scorpions probably contributed to the high diversity and endemism of Parabuthus and Opistophthalmus, and may, in part, explain the larger number of hotspots in the arid, sandy western half of southern Africa.

Pelophilous species are capable of utilising a greater range of substratum hardness than other fossorial scorpions (Lamoral, 1978a). Accordingly, they may be less easily isolated and less prone to speciation. The wide distributions of several pelophilous species (e.g. *O. glabrifrons*) contrast markedly with the often highly restricted distributions of most lithophilous and psammophilous species. Nevertheless, many pelophilous species of *Opistophthalmus* also display markedly discrete distributions. Such species tend to be concentrated in the mountainous regions of Namibia and the Northern, Western, and Eastern Cape provinces of South Africa, related species invariably occupying adjacent, but separate mountain ranges, or the intervening valleys. This is particularly clear among semi-lithophilous species of *Opistophthalmus* (Prendini, 2001a). The adults of such species construct shallow scrapes under rocks (Eastwood, 1978b), and may have speciated in a manner similar to that proposed for the lithophilous

*Hadogenes*, where the ecological requirement for rock cover promoted speciation by vicariance in the various discrete mountain ranges.

## 4.3.4 Drainage patterns

The most important agents of vicariance in the evolution of southern African scorpions, besides the sand systems and mountain ranges, are represented by major rivers, especially the Orange, the largest river crossing the arid interior and western coastline of southern Africa. The scorpion faunas occurring to the north and south of this river are distinctly different, sharing only 20 species that occur on both sides. All remaining species are endemic to the region north or south of the river and, given their low vagility, are probably autochthonous. The present disjunction is so complete that although several species can be found right up to the northern and southern banks of the river, they do not occur on the opposite sides, a pattern mirrored by flightless tenebrionid and scarabaeid beetles, and lepismatid silverfish (Penrith 1975, 1977; Irish 1990; Harrison 1999).

Other rivers that appear to have acted as agents of vicariance in the evolution of southern African scorpions include the Curoca, Kunene, Hoanib, Huab, Ugab, Swakop, Kuiseb, Koichab, Olifants and Berg rivers of the west coast, the Fish River, a major tributary of the Orange in the southern interior of Namibia, the Breë River of the south coast. and the Limpopo and Zambezi rivers of the east coast. In addition to the Limpopo and the Zambezi, the presence of numerous large, fast-flowing rivers emanating from the Natal Drakensberg section of the eastern escarpment might explain the absence of Opistophthalmus species from the region of South Africa between the Natal Drakensberg, the Kei River (Eastern Cape Province) and the Tugela River (KwaZulu-Natal Province). The Kei and Tugela rivers have both incised deep valleys that are inhabited by endemic species of Opistophthalmus, but no Opistophthalmus species are found in between them. The effects of river drainage on the evolution and biogeography of southern African scorpions are currently under further investigation by the author.

## 5. CONCLUSIONS

The southern African scorpion fauna contains the following components: ancient elements comprising endemic Gondwanaland relicts (*Brandbergia*, *Lisposoma*, *Karasbergia*, *Opisthacanthus*) and their endemic southern African derivatives (*Cheloctonus*); old elements now widespread in the Afrotropical and Oriental regions (*Hottentotta*, *Lychas* and *Uroplectes*),

many of which are also endemic to the southern African subregion, and their endemic derivatives (*Afroisometrus*, *Pseudolychas*); recent elements derived in and largely endemic to the Afrotropical region (*Parabuthus*) or the southern African subregion (*Hadogenes* and *Opistophthalmus*).

Combined effects of geomorphology and palaeoclimatic change have acted as agents of vicariance in determining the evolution of the southern African scorpions, with their restricted climatic and substratal requirements. and are primarily responsible for the high species richness and endemism in the subregion (Prendini, 2001a). The progressive spatial restriction of groups of taxa, such as an entire genus, groups of related species, or single species, is the result of vicariance, in turn facilitated by limited vagility due to narrow ecological tolerances. In many cases, agents of vicariance are still in existence (e.g. mountain ranges, sand systems and palaeodrainage channels), and coincide with disjunct distribution patterns. Specialist lithophilous (Hadogenes), semi-psammophilous or psammophilous (Parabuthus and Opistophthalmus), and pelophilous (Opistophthalmus) taxa have speciated extensively, producing a high proportion of range-restricted endemics. Opistophthalmus, in particular, with 42 % of the southern African species, has radiated in a similar manner to the burrowing scorpionoid genus Urodacus in Australia (Koch, 1977, 1978, 1981). Presumably specialisation into psammophilous, pelophilous, and semi-lithophilous ecomorphotypes promoted rampant speciation by vicariance in both genera (Prendini, 2001a). This pattern contrasts with that observed in the relictual genera (e.g., Lisposoma, Karasbergia, Pseudolychas, Cheloctonus, and Opisthacanthus), which contain relatively few species.

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#### REFERENCES

- Aguiar, O. B., 1978, Alguns escorpiões de Moçambique, Garcia de Orta, Sér. Zool., Jta. Investig. Cien. Ultramar 7:107-114.
- Armas, L. F., de, 1981, Primero hallazgos de la familia Diplocentridae (Arachnida: Scorpionida) en la Española, *Poeyana* 213:1-12.
- Armas, L. F., de, 1984, Escorpiones del Archipiélago Cubano. 7. Adiciones y enmiendas (Scorpiones: Buthidae, Diplocentridae), *Poeyana* 275:1-37.
- Armas, L. F., de, 2001, Scorpions of the Greater Antilles, with the description of a new troglobitic species (Scorpiones: Diplocentridae), in: Scorpions 2001. In Memoriam Gary A. Polis, V. Fet and P. A. Selden, eds., British Arachnological Society, Burnham Beeches, Bucks, UK, pp. 245-253.
- Armas, L. F., de, and Marcano Fondeur, E. de J., 1987, Nuevos escorpiones (Arachnida: Scorpiones) de República Dominicana, *Poeyana* 356:1-24.
- Balinsky, B. I., 1962, Patterns of animal distribution on the African continent, Ann. Cape Prov. Mus. 2:299-309.
- Bamps, P., 1982, Flora d'Afrique Centrale (Zaïre-Rwanda-Burundi). Répertoire de lieux de récolte, Jardin botanique national de Belgique, Meise.
- Barnard, P., 1998, *Biological Diversity in Namibia*: A *Country Study*, Namibian National Biodiversity Task Force, Windhoek.
- Byalynitskii-Birulya, A. A. [Birula, A. A.], 1917a, Arachnoidea Arthrogastra Caucasica. Pars I. Scorpiones, Zapiski Kavkazskogo Muzeya (Mémoires du Musée du Caucase), Imprimerie de la Chancellerie du Comité pour la Transcaucasie, Tiflis, Russia. Ser A, 5:1-253. [Russian; English translation: Byalynitskii-Birulya, A. A., 1964, Arthrogastric Arachnids of Caucasia. 1. Scorpions, Israel Program for Scientific Translations, Jerusalem].
- Byalynitskii-Birulya, A. A. [Birula, A. A.], 1917b, Faune de la Russie et des pays limitrophes fondee principalement sur les collections du Musée zoologique de l'Académie des sciences de Russie. Arachnides (Arachnoidea), 1:1-227. [Russian, English translation: Byalynitskii-Birulya, A. A., 1965, Fauna of Russia and adjacent countries. Arachnoidea. Vol. I. Scorpions, Israel Program for Scientific Translations, Jerusalem].
- Birula, A. A., 1925, Skorpiologische Beiträge. 10. Zur geographischen Verbreitung zweier weitverbreiteter Skorpionen-Arten Palaearcticums, *Zool. Anz.* 63:93-96.
- Bradley, R., 1986, The relationship between population density of *Paruroctonus utahensis* (Scorpionida: Vaejovidae) and characteristics of its habitat, *J. Arid Environ.* 11:165-172.
- Bradley, R., 1988, The behavioural ecology of scorpions—a review, in: *Australian Arachnology. Australian Entomological Society Miscellaneous Publication* 5, A. D. Austin and N. W. Heather, eds., Watson, Ferguson and Co., Brisbane, pp. 23-36.
- Bradley, R., and Brody, A., 1984, Relative abundance of three vaejovid scorpions across a habitat gradient, *J. Arachnol.* 11:437-440.
- Coaton, W. G. H., and Sheasby, J. L. 1972, Preliminary report on a survey of the termites (Isoptera) of South West Africa, *Cimbebasia Memoir* 2:1-129.
- Copeland, D. J. B., 1966, Gazetteer of geographical names in the Republic of Zambia, Government Printer, Lusaka.
- Davis, D. H. S., and Misonne, X., 1964, Gazetteer of collecting localities of African rodents, Documentation Zoologique (Tervuren) 7:1-100.
- De Meillon, B., Davis, D. H. S., and Hardy, F., 1961, *Plague in southern Africa*, Vol 1, Government Printer, Pretoria.
- Doidge, E. M., 1950, The South African fungi and lichens to the end of 1945. Locality index, Bothalia 5:967-994.

- Eastwood, E. B., 1977a, Notes on the scorpion fauna of the Cape. Part 1. Description of neotype of Opisthophthalmus capensis (Herbst) and remarks on the O. capensis and O. granifrons Pocock species-groups (Arachnida, Scorpionida, Scorpionidae), Ann. S. Afr. Mus. 72:211-226.
- Eastwood, E. B., 1977b, Notes on the scorpion fauna of the Cape. Part 2. The *Parabuthus capensis* (Ehrenberg) species-group; remarks on taxonomy and bionomics (Arachnida, Scorpionida, Buthidae), *Ann. S. Afr. Mus.* 73:199-214.
- Eastwood, E. B., 1978a, Notes on the scorpion fauna of the Cape. Part 3. Some observations on the distribution and biology of scorpions on Table Mountain, Ann. S. Afr. Mus. 74:229-248.
- Eastwood, E. B., 1978b, Notes on the scorpion fauna of the Cape. Part 4. The burrowing activities of some scorpionids and buthids (Arachnida, Scorpionida), Ann. S. Afr. Mus. 74:249-255.
- Endrödy-Younga, S., 1982, Dispersion and translocation of dune specialist tenebrionids in the Namib area, Cimbebasia (A) 5:257-271.
- Fet, V., 2000a, Family Scorpionidae Latreille, 1802, in: Catalog of the Scorpions of the World (1758-1998), V. Fet, W. D. Sissom, G. Lowe and M. E. Braunwalder, The New York Entomological Society, New York, pp. 427-486.
- Fet, V., 2000b, Family Ischnuridae Simon, 1879, in: Catalog of the Scorpions of the World (1758-1998), V. Fet, W. D. Sissom, G. Lowe and M. E. Braunwalder, The New York Entomological Society, New York, pp. 383-408.
- Fet, V., and Lowe, G., 2000, Family Buthidae C. L. Koch, 1837, in: *Catalog of the Scorpions of the World (1758-1998)*, V. Fet, W. D. Sissom, G. Lowe and M. E. Braunwalder, The New York Entomological Society, New York, pp. 54-286.
- Fet, V., Polis, G. A., and Sissom, W. D., 1998, Life in sandy deserts: The scorpion model, J. Arid Environ. 39:609-622.
- Fet, V., Capes, M., and Sissom, W. D., 2001, A new genus and species of psammophilic scorpion from eastern Iran (Scorpiones: Buthidae), in: *Scorpions 2001*. In Memoriam *Gary A. Polis*, V. Fet and P. A. Selden, eds., British Arachnological Society, Burnham Beeches, Bucks, UK, pp. 183-189.
- FitzPatrick, M. J., 1994a, A new species of Lychas C.L. Koch, 1845 from Zimbabwe (Scorpionida: Buthidae), Arnoldia Zimbabwe 10:23-28.
- FitzPatrick, M. J., 1994b, A checklist of the *Parabuthus* Pocock species of Zimbabwe with a re-description of *P. mossambicensis* (Peters, 1861) (Arachnida: Scorpionida), *Trans. Zimbabwe Sci. Assoc.* 68:7-14.
- FitzPatrick, M. J., 1996, The genus *Uroplectes* Peters, 1861 in Zimbabwe (Scorpiones: Buthidae), *Arnoldia Zimbabwe* 10:47-70.
- FitzPatrick, M. J., 2001, Synonymy of some *Uroplectes* Peters, 1861 (Scorpiones: Buthidae), in: *Scorpions 2001*. In Memoriam *Gary A. Polis*, V. Fet and P. A. Selden, eds., British Arachnological Society, Burnham Beeches, Bucks, UK, pp. 191-193.
- Francke, O. F., 1974, Nota sobre los generos *Opisthacanthus* Peters y *Nepabellus* nom. nov. (Scorpionida, Scorpionidae) e informe sobre el hallazgo de *O. lepturus* en la isla del Coco, Costa Rica, *Brenesia* 4:31-35.
- Francke, O. F., 1978, Systematic revision of diplocentrid scorpions from circum-Caribbean lands, Spec. Publ. Texas Tech Univ. 14:1-92.
- Francke, O. F., 1979, Additional record of *Heteronebo* from Abd-el-Kuri Island, P.D.R. Yemen (Scorpiones: Diplocentridae), *J. Arachnol.* 7:265.
- Francke, O. F., 1982, Are there any bothriurids (Arachnida, Scorpiones) in southern Africa?, J. Arachnol. 10:35-39.
- Francke, O. F., 1985, Conspectus genericus scorpionum, 1758-1982 (Arachnida, Scorpiones), Occas. Pap. Mus., Texas Tech Univ. 98:1-32.

- Francke, O. F., and Sissom, W. D., 1980, Scorpions from the Virgin Islands (Arachnida, Scorpiones), Occas. Pap. Mus., Texas Tech Univ. 65:1-19.
- Gonçalves, M. L., 1962, Índice Toponímico de Moçambique, Centro de Botânica de Junta de Investigações do Ultramar, Lisboa.
- González-Sponga, M. A., 1972, Ananteris venezuelensis (Scorpionida, Buthidae) nueva especie de la Guayana de Venezuela, Mem. Soc. Cienc. Nat. "La Salle" 32:205-214.
- González-Sponga, M. A., 1978, Escorpiofauna de la Región Oriental del Estado Bolivar, Venezuela. Ed. Roto-Impresos, Caracas.
- González-Sponga, M. A., 1980, *Ananteris turumbanensis*, nueva especie de la Guayana de Venezuela (Scorpionida, Buthidae), *Mem. Soc. Cienc. Nat.* "La Salle" **40**:95-107.
- González-Sponga, M. A., 1984, Escorpiones de Venezuela, Cuadernos Lagoven, Caracas.
- González-Sponga, M. A., 1996, *Guía para identificar escorpiones de Venezuela*, Cuadernos Lagoven, Caracas.
- Gross, A., 1920, Gazetteer of Central and South Africa, Geographia 1-271.
- Gromov, A.V., 2001. The northern boundary of scorpions in Central Asia, in: *Scorpions* 2001. In Memoriam *Gary A. Polis*, V. Fet and P. A. Selden, eds., British Arachnological Society, Burnham Beeches, Bucks, UK, pp. 301-306.
- Haacke, W. D., 1975, The burrowing geckos of southern Africa, 1 (Reptilia: Gekkonidae), Ann. Transv. Mus. 29:197–241.
- Harington, A., 1978, Burrowing biology of the scorpion *Cheloctomus jonesii* Pocock (Arachnida: Scorpionida: Scorpionidae), *J. Arachnol.* 5:243-249.
- Harington, A., 1984, Character variation in the scorpion *Parabuthus villosus* (Peters) (Scorpiones, Buthidae): a case of intermediate zones, *J. Arachnol.* 11:393-406.
- Harington, A., 2001 [2002]. Description of a new species of *Opisthophthalmus* C.L. Koch (Scorpiones, Scorpionidae) from southern Namibia. *Rev. arachnol.* 14:25-30.
- Harrison, J. du G., 1999, Systematics of the endemic south-west African dung beetle genus *Pachysoma* MacLeay (Scarabaeidae: Scarabaeinae), M.Sc. Thesis, University of Pretoria.
- Haydon, D. T., Crother, B. I., and Pianka, E. R., 1994, New directions in biogeography?, Trends Ecol. Evol. 9:403-406.
- Hewitt, J., 1918, A survey of the scorpion fauna of South Africa, *Trans. R. Soc. S. Afr.* 6:89-192.
- Hewitt, J., 1925, Facts and theories on the distribution of scorpions in South Africa, Trans. R. Soc. S. Afr. 12:249-276.
- Irish, J., 1988, Gazetteer of place names on maps of Botswana, Cimbebasia 10:107-146.
- Irish, J., 1990, Namib biogeography, as exemplified mainly by the Lepismatidae (Thysanura: Insecta), in: *Namib Ecology: 25 Years of Namib Research, Transvaal Museum Monograph* 7, M. K. Seely, ed., Transvaal Museum, Pretoria, pp. 61-66.
- Jeram, A., 1990a, Book-lungs in a Lower Carboniferous scorpion, Nature 343:360-361.
- Jeram, A., 1990b, When scorpions ruled the world, New Scientist 126:52-55.
- Jeram, A., 1993 [1994a], Scorpions from the Viséan of East Kirkton, West Lothian, Scotland, with a revision of the infraorder Mesoscorpionina, Trans. R. Soc. Edinburgh: Earth Sci. 84:283-299.
- Jeram, A., 1994b, Carboniferous Orthosterni and their relationship to living scorpions, Palaeontology 37:513-550.
- Keast, A., 1981, Ecological biogeography of Australia, Junk, The Hague.
- Koch, L. E., 1977, The taxonomy, geographic distribution and evolutionary radiation of Australo-Papuan scorpions, Rec. West. Aust. Mus. 5:83-367.
- Koch, L. E., 1978, A comparative study of the structure, function and adaptation to different habitats of burrows in the scorpion genus *Urodacus* (Scorpionida, Scorpionidae), *Rec. West. Aust. Mus.* 6:119-146.
- Koch, L. E., 1981, The scorpions of Australia: aspects of their ecology and zoogeography, in: Ecological biogeography of Australia, A. Keast, ed., Junk, The Hague, pp. 875-884.

- Kraepelin, K., 1905, Die geographische Verbreitung der Skorpione. Zool. Jahrb., Abt. Syst. 22:321-364.
- Kraepelin, K. 1912 [1913]. Neue Beiträge zur Systematik der Gliederspinnen. III. A. Bemerkungen zur Skorpionenfauna Indiens. B. Die Skorpione, Pedipalpen und Solifugen Deutsch-Ost-Afrikas, Jahrb. Hamburg Wiss. Anst. 30:123-196.
- Lamoral, B. H., 1978a, Soil hardness, an important and limiting factor in burrowing scorpions of the genus *Opisthophthalmus* C.L. Koch, 1837 (Scorpionidae, Scorpionida), *Symp. zool. Soc. Lond.* (London) 42:171-181.
- Lamoral, B. H., 1978b, The scorpions of South West Africa. Ph.D. thesis, University of Natal, Pietermaritzburg.
- Lamoral, B. H., 1979, The scorpions of Namibia (Arachnida: Scorpionida), *Ann. Natal Mus.* 23:497-784.
- Lamoral, B. H., 1980a, A reappraisal of the suprageneric classification of Recent scorpions and of their zoogeography, in: Verhandlungen 8. Internationaler Arachnologen-Kongress Abgehalten an der Universität für Bodenkultur Wien, 7-12 Juli, 1980, J. Grüber, ed., H. Egermann, Vienna, pp. 439-444.
- Lamoral, B. H., 1980b, Two new psammophile species and new records of scorpions from the northern Cape Province of South Africa (Arachnida: Scorpionida), Ann. Natal Mus. 24:201-210.
- Lamoral, B. H., and Reynders, S. C., 1975, A catalogue of the scorpions described from the Ethiopian Faunal Region up to December 1973, Ann. Natal Mus. 22:489-576.
- Lancaster, N., 1990, Regional aeolian dynamics in the Namib, in: Namib Ecology: 25 Years of Namib Research, Transvaal Museum Monograph 7, M. K. Seely, ed., Transvaal Museum, Pretoria, pp. 39-46.
- Lawrence, R. F., 1942, The scorpions of Natal and Zululand, Ann. Natal Mus. 10:221-235.
- Lawrence, R. F., 1952, The unequal distribution of some invertebrate animals in South Africa. S. Afr. J. Sci. 48:308-310.
- Lawrence, R. F., 1953, The Biology of the Cryptic Fauna of Forests, with Special Reference to the Indigenous Forest of South Africa. A.A. Balkema, Cape Town.
- Lawrence, R. F., 1955, Solifugae, scorpions and Pedipalpi, with checklists and keys to South African families, genera and species. Results of the Lund University Expedition in 1950– 1951, in: South African Animal Life, Vol. 1, B. Hanström, P. Brinck and G. Rudebeck, eds., Almqvist & Wiksells, Uppsala, pp. 152-262.
- Lawrence, R. F., 1969, A new genus of psammophile scorpion and new species of Opisthophthalmus from the Namib Desert, Sci. Pap. Namib Desert Res. stn 48:105-116.
- Leistner, O. A., and Morris, J. W., 1976, Southern African place names, *Ann. Cape Prov. Mus.* 12:1-565.
- Lombard, A. T., 1995a, Introduction to an evaluation of the protection status of South Africa's vertebrates, S. Afr. J. Zool. 30:63-70.
- Lombard, A. T., 1995b, The problems with multi-species conservation: Do hotspots, ideal reserves and existing reserves coincide?, S. Afr. J. Zool. 30:145-163.
- Lourenço, W. R., 1980, A propósito de duas novas espécies de Opisthacanthus para a região Neotropical, Opisthacanthus valerioi da "Isla del Coco", Costa Rica e Opisthacanthus heurtaultae da Guiana Francesa (Scorpiones, Scorpionidae), Rev. Nordest. Biol. 3:179-194.
- Lourenço, W. R., 1981, Opisthacanthus lamorali, nouvelle espèce de Scorpionidae pour la Région Afrotropicale (Arachnida: Scorpionidae), Ann. Natal Mus. 24:625-634.
- Lourenço, W. R., 1982, Révision du genre *Ananteris* Thorell, 1891 (Scorpiones, Buthidae) et description de six espèces nouvelles, *Bull. Mus. natn. Hist. nat.*, *Paris*, *4e sér.* 4:119-151.
- Lourenço, W. R., 1983a, Le faune des scorpiones de Guyane française, Bull. Mus. natn. Hist. nat., Paris, 4e sér. 5:771-808.

- Lourenço, W. R., 1983b, Contributions à la connaissance de la faune hypsophile du Malawi (Mission R. Jocque). 2. Scorpions, *Rev. Zool. Afr.* 97:192-201.
- Lourenço, W. R., 1985, Le vériyable statut des genres *Ananteris* Thorell, 1891 et *Ananteroides* Borelli, 1911 (Scorpiones, Buthidae), *Ann. Natal Mus.* 26:407-416.
- Lourenço, W. R., 1984a, La biogéographie des scorpions sud-américains (problèmes et perspectives), Spixiana 7:11-18.
- Lourenço, W. R., 1984b, Ananteris luciae, nouvelle espèce de scorpion de l'Amazonie brasilienne (Scorpiones, Buthidae), J. Arachnol. 12:279-282.
- Lourenço, W. R., 1987a, Révision systématique des scorpions du genre *Opisthacanthus* (Scorpiones, Ischnuridae), *Bull. Mus. natn. Hist. nat.*, *Paris*, *4e sér.* 9:887-931.
- Lourenço, W. R., 1987b, Déscription d'une nouvelle espèce d'Ananteris collectee dans l'état de Maranhão, Brésil (Scorpiones, Buthidae), Bol. Mus. Para. Emilio Goeldi, Sér. Zool. 3:19-23.
- Lourenço, W. R., 1988, Considérations biogéographiques, écologiques et évolutives sur les espèces néotropicales d'Opisthacanthus Peters, 1861 (Scorpiones, Ischnuridae), Stud. Neotrop. Fauna Envir. 23:41-53.
- Lourenço, W. R., 1989, Rétablissement de la famille des Ischnuridae distincte des Scorpionidae Pocock, 1893, Rev. Arachnol. 8:159-177.
- Lourenço, W. R., 1991a, *Opisthacanthus*, genre Gondwanien défini comme groupe naturel. Caractérisation des sous-genres et des groupes d'espèces (Arachnida, Scorpiones, Ischnuridae), *Iheringia*, *Sèr. Zool.* 71:5-42.
- Lourenço, W. R., 1991b, Scorpion species biodiversity in tropical South America and its application in conservation programmes, Amer. Zool. 31:A85-A85.
- Lourenço, W. R., 1990 [1991c], Les scorpions (Chelicerata) de Colombie. II. Les faunes des régions de Santa Marta et de la Cordillère orientale. Approche biogéographique, Senkenbergiana Biol. 71:275-288.
- Lourenço, W. R., 1993, A review of the geographical distribution of the genus *Ananteris* Thorell (Scorpiones: Buthidae), with description of a new species, *Rev. Biol. Trop.* 41:697-701.
- Lourenço, W. R., 1994a, Scorpion biogeographic patterns as evidence for a Neblina-São Gabriel endemic center in Brazilian Amazonia, *Rev. Acad. Colomb. Cienc.* **19**:181-185.
- Lourenço, W. R., 1994b, Biogeographic patterns of tropical South American scorpions, *Stud. Neotr. Fauna Environ.* **29**:219-231.
- Lourenço, W. R., 1994c, Diversity and endemism in tropical versus temperate scorpion communities, Biogeographica 70:155-160.
- Lourenço, W. R., 1995, Nouvelles considérations sur la classification et la biogéographie des *Opisthacanthus* néotropicaux (Scorpiones, Ischnuridae), *Biogeographica* 71:75-82.
- Lourenço, W. R., 1996, The biogeography of scorpions, Rev. Suisse Zool., Vol. hors série II:437-448 [Proceedings of the XIIIth International Congress of Arachnology, Geneva, 3-8 September 1995, V. Mahnert, ed.].
- Lourenço, W. R., 1997, Synopsis de la faune de scorpions de Colombie, avec des considérations sur la systématique et la biogéographie des espèces, Rev. Suisse Zool. 104:61-94.
- Lourenço, W. R., 1998, Panbiogéographie, les distributions disjontes et le concept de familie relictuelle chez les scorpions, *Biogeographica* 74:133-144.
- Lourenço, W. R., 1999, Considérations taxonomiques sur le genre Hadogenes Kraepelin, 1894; création de la sous-famille des Hadogeninae n. subfam., et description d'une espèce nouvelle pour l'Angola (Scorpiones, Scorpionidae, Hadogeninae), Rev. Suisse Zool. 106:929-938.
- Lourenço, W. R., 2000, Panbiogéographie, les familles des scorpions et leur répartition géographique, Biogeographica 76:21-39.

- Lourenço, W. R., 2001, Nouvelles considerations sur la phylogenie et la biogéographie des scorpions Ischnuridae de Madagascar, Biogeographica 77:83-96.
- Lourenço, W. R., and Fé, N. F., 2003, Description of a new species of *Opisthacanthus* Peters (Scorpiones, Liochelidae) to Brazilian Amazonia, *Rev. Iber. Aracnol.* **8**:81-88.
- Lourenço, W. R., and Flórez, E., 1989, Los escorpiones (Chelicerata) de Colombia. I. La Fauna de la Isla Gorgona. Approximación biogeográfica, *Caldasia* 16:66-70.
- MacArthur, R. H., 1972, Geographical ecology, Harper and Row, New York.
- Main, B. Y., 1956, Taxonomy and biology of the genus Isometroides Keyserling (Scorpionida), Aust. J. Zool. 4:158-164.
- Mayr, E., 1942, Systematics and the Origin of Species, Columbia University Press, New York.
- Mayr, E., 1963, Animal Species and Evolution, Harvard University Press, Cambridge, MA.
- McCormick, S. J., and Polis, G. A., 1990, Prey, predators, parasites, in: The Biology of Scorpions, G. A. Polis, ed., Stanford University Press, Stanford, CA, pp. 294-320.
- Midgley, D. C., Pitman, W. V., and Middleton, B. J., 1994, Surface Water Resources of South Africa 1990 (WR90), CD-ROM, Water Research Commission, Pretoria.
- Ministry of Land and Natural Resources (Northern Rhodesia), 1959, Gazetteer of Geographical Names in the Barotseland Protectorate, Government Printer, Lusaka.
- Ministry of Land and Mines (Zambia), 1967, Gazetteer of Names in the Republic of Zambia, Government Printer, Lusaka.
- Mugo, D. N., Lombard, A. T., Bronner, G. N., Gelderblom, C. M., and Benn, G. A., 1995, Distribution and protection of endemic or threatened rodents, lagomorphs and macrosceledids in South Africa, S. Afr. J. Zool. 30:115-126.
- Myers, A. A., and Giller, P. S., 1988, Process, pattern and scale in biogeography, in:

  Analytical Biogeography: An Integrated Approach to the Study of Animal and Plant Distributions, A. A. Myers and P. S. Giller, eds., Chapman and Hall, London, pp. 3-21.
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A. B., and Kent, J., 2000, Biodiversity hotspots for conservation priorities, *Nature* 403:853-858.
- National Place Names Committee (Department of National Education, Republic of South Africa), 1951, *Official Place Names in the Union and South West Africa* (Approved to end 1948), Government Printer, Pretoria.
- National Place Names Committee (Department of National Education, Republic of South Africa), 1978, Official Place Names in the Republic of South Africa and South West Africa (Approved to 1 April 1977), Government Printer, Pretoria.
- National Place Names Committee (Department of National Education, Republic of South Africa), 1988, *Official Place Names in the Republic of South Africa* (Approved to 1977 to 1988), Government Printer, Pretoria.
- Nenilin, A. B., and Fet, V., 1992, [Zoogeographical analysis of the world scorpion fauna (Arachnida: Scorpiones)], Arthopoda Selecta, Moscow 1:3-31. [Russian].
- Newlands, G., 1969, Two new scorpions from the northern Transvaal, J. ent. Soc. sth. Afr. 32:5-7.
- Newlands, G., 1972a, Notes on psammophilous scorpions and a description of a new species (Arachnida: Scorpionida), *Ann. Transv. Mus.* 27:241-257.
- Newlands, G., 1972b, Ecological adaptations of Kruger National Park scorpionids (Arachnida: Scorpiones), *Koedoe* 15:37-48.
- Newlands, G., 1972c, A description of *Hadogenes lawrencei* sp. nov. (Scorpiones) with a checklist and key to the South-West African species of the genus *Hadogenes*, *Madoqua*, ser. 2. 1:133-140.
- Newlands, G., 1973, Zoogeographical factors involved in the trans-Atlantic dispersal pattern of the genus *Opisthacanthus* Peters (Arachnida: Scorpionidae), *Ann. Transv. Mus.* 28:91-100.
- Newlands, G., 1974, Transvaal scorpions, Fauna and Flora 25:3-7.

- Newlands, G., 1978a, Arachnida (except Acari), in: *Biogeography and Ecology of southern Africa*, Vol. 2, M. J. A. Werger, ed., Junk, The Hague, pp. 677-684.
- Newlands, G., 1978b, Review of southern African scorpions and scorpionism, S. Afr. Med. J. 54:613-615.
- Newlands, G., 1980, A revision of the scorpion genus *Hadogenes* Kraepelin 1894 (Arachnida: Scorpionidae) with a checklist and key to the species, M.Sc. thesis, Potchefstroom University for C.H.E.
- Newlands, G., and Cantrell, A. C., 1985, A re-appraisal of the rock scorpions (Scorpionidae: Hadogenes), Koedoe 28:35-45.
- Newlands, G., and Martindale, C. B., 1980, The buthid scorpion fauna of Zimbabwe-Rhodesia with checklists and keys to the genera and species, distributions and medical importance (Arachnida: Scorpiones), Z. Angew. Zool. 67:51-77.
- Newlands, G., and Prendini, L., 1997, Redescription of *Hadogenes zumpti* (Scorpiones: Ischnuridae), an unusual rock scorpion from the Richtersveld, South Africa, S. Afr. J. Zool. 32:76-81.
- Penrith, M.-L., 1975, The species of *Onymacris* (Coleoptera: Tenebrionidae), *Cimbebasia* (A) 4:47-97.
- Penrith, M.-L., 1977, The Zophosini (Coleoptera: Tenebrionidae) of western southern Africa, Cimbebasia Memoir 3:1-291.
- Penrith, M.-L., 1979, Revision of the western southern African Adesmiini (Coleoptera: Tenebrionidae), *Cimbebasia* (A) 5:1-94.
- Penrith, M.-L., 1981a, Revision of the Zophosini (Coleoptera: Tenebrionidae). Part 2. The subgenus *Zophosis* Latreille, and seven related south-western African subgenera, *Cimbebasia* (A) **6**:17-109.
- Penrith, M.-L., 1981b, Revision of the Zophosini (Coleoptera: Tenebrionidae). Part 4. Twelve subgenera from arid southern Africa, *Cimbebasia* (A) 6:125-164.
- Penrith, M.-L., 1987, Revision of the genus *Tarsocnodes* Gebien (Coleoptera: Tenebrionidae: Molurini), and a description of a monotypical genus from the Kalahari, *Cimbebasia* (A) 7:235-270.
- Platnick, N. I., and Nelson, G., 1978, A method of analysis for historical biogeography, Syst. Zool. 27:1-16.
- Pocock, R. I., 1894, Scorpions and their geographical distribution, Nat. Sci. 4:353-364.
- Polhill, D., 1988, Flora of Tropical East Africa. Index of Collecting Localities, Royal Botanic Gardens, Kew.
- Polis, G. A., 1979, Prey and feeding phenology of the desert sand scorpion *Paruroctonus mesaensis* (Scorpionida: Vaejovidae), *J. Zool.* (Lond.) 188:333-346.
- Polis, G. A., 1990. Ecology, in: The Biology of Scorpions, G. A. Polis, ed., Stanford University Press, Stanford, CA, pp. 247-293.
- Polis, G. A., and Holt, R. D., 1992, Intraguild predation: the dynamics of complex trophic interactions, *Trends Ecol. Evol.* 7:151-154.
- Polis, G. A., and McCormick, S. J., 1986, Patterns of resource use and age structure among a guild of desert scorpions, *J. Anim. Ecol.* **55**:59-73.
- Polis, G. A., and McCormick, S. J., 1987, Intraguild predation and competition among desert scorpions, *Ecology* 68:332-343.
- Polis, G. A., Sissom, W. D., and McCormick, S. J., 1981, Predators of scorpions: field data and a review, *J. Arid Environ.* 4:309-326.
- Postmaster-General (Republic of South Africa), 1970, List of Post Offices in the Republic of South Africa, in South-West Africa, and other countries of the African Postal Union, Government Printer, Pretoria.
- Poynton, J. C., 1964, The Amphibia of southern Africa: a faunal study, Ann. Natal Mus. 17:1-334.

- Poynton, J. C., and Broadley, D. G., 1985a, Amphibia Zambesiaca 1. Scolecomorphidae, Pipidae, Microhylidae, Hemisidae, Arthroleptidae, Ann. Natal Mus. 26:503-553.
- Poynton, J. C., and Broadley, D. G., 1985b. Amphibia Zambesiaca 2. Ranidae, *Ann. Natal Mus.* 27:115-181.
- Poynton, J. C., and Broadley, D. G., 1987, Amphibia Zambesiaca 3. Rhacophoridae and Hyperoliidae, Ann. Natal Mus. 28:161-229.
- Prendini, L., 2000a, Phylogeny and classification of the superfamily Scorpionoidea Latreille 1802 (Chelicerata, Scorpiones): An exemplar approach, *Cladistics* 16:1-78.
- Prendini, L., 2000b, A new species of *Parabuthus* Pocock (Scorpiones, Buthidae), and new records of *Parabuthus capensis* (Ehrenberg), from Namibia and South Africa, *Cimbebasia* 16:201-214.
- Prendini, L., 2000c, Chelicerata (Scorpiones), in: Dâures Biodiversity of the Brandberg Massif, Namibia, A.H. Kirk-Spriggs and E. Marais, eds., Cimbebasia Memoir 9:109-120.
- Prendini, L., 2001a, Substratum specialization and speciation in southern African scorpions: the Effect Hypothesis revisited, in: Scorpions 2001. In Memoriam Gary A. Polis, V. Fet and P. A. Selden, eds., British Arachnological Society, Burnham Beeches, Bucks, UK, pp. 113-138.
- Prendini, L., 2001b, Phylogeny of Parabuthus (Scorpiones, Buthidae), Zool. Scr. 30:13-35.
- Prendini, L., 2001c, Two new species of *Hadogenes* (Scorpiones, Ischnuridae) from South Africa, with a redescription of *Hadogenes bicolor* and a discussion on the phylogenetic position of *Hadogenes*, *J. Arachnol.* 29:146-172.
- Prendini, L., 2001d, A review of synonyms and subspecies in the genus *Opistophthalmus C.L.* Koch (Scorpiones: Scorpionidae), *Afr. Entomol.* 9:17-48.
- Prendini, L., 2003a, A new genus and species of bothriurid scorpion from the Brandberg Massif, Namibia, with a reanalysis of bothriurid phylogeny and a discussion of the phylogenetic position of *Lisposoma* Lawrence, *Syst. Ent.* 28:149-172.
- Prendini, L., 2003b, Revision of the genus *Lisposoma* Lawrence, 1928 (Scorpiones: Bothriuridae), *Insect Syst. Evol.* 34:241-264.
- Prendini, L., 2003c, Discovery of the male of *Parabuthus muelleri*, and implications for the phylogeny of *Parabuthus* (Scorpiones: Buthidae), *Amer. Mus. Novit.* **3408**:1-24.
- Prendini, L., 2004a, The systematics of southern African *Parabuthus* Pocock (Scorpiones, Buthidae): Revisions to the taxonomy and key to the species, *J. Arachnol.* **32**:109-186.
- Prendini, L., 2004b, Systematics of the genus *Pseudolychas* Kraepelin (Scorpiones: Buthidae), *Ann. Entomol. Soc. Amer.* 97:37-63.
- Prendini, L., in press, On *Hadogenes angolensis* Lourenço, 1999 syn. n. (Scorpiones, Liochelidae), with a redescription of *H. taeniurus* (Thorell, 1876). *Rev. Suisse Zool*.
- Prendini, L., in press, Revision of *Karasbergia* Hewitt (Scorpiones: Buthidae), a monotypic genus endemic to southern Africa, *J. Afrotrop. Zool*.
- Prendini, L., Crowe, T. M., and Wheeler, W. C., 2003, Systematics and biogeography of the family Scorpionidae Latreille, with a discussion of phylogenetic methods, *Invert. Syst.* 17:185-259.
- Raffle, J. A., 1984, Third report of the Place Names Commission, Government Printer, Gaborone.
- Raper, P. E., 1987, Dictionary of Southern African Place Names, Lowry Publishers, Johannesburg.
- San Martín, P. R., 1961, Observaciones sobre la ecología y distribución geográfica de tres especes de escorpiones en el Uruguay. Rev. Fac. Human. Cien., Univ. Repúb. (Montevideo) 19:1-42.
- Selden, P. A., 1993, Fossil arachnids—recent advances and future prospects, Mem. Queensl. Mus. 33:389-400.

- Shorthouse, D. J., and Marples, T., 1980, Observations on the burrow and associated behaviour of the arid-zone scorpion *Urodacus yaschenkoi* (Birula), *Aust. J. Zool.* **28**:581-590.
- Simberloff, D., 1983, Sizes of coexisting species, in: *Coevolution*, D.J. Futuyma and M. Slatkin, eds., Sinauer Associates, Sunderland, MA, pp. 404-430.
- Sissom, W. D., 1990, Systematics, biogeography and palaeontology, in: *The Biology of Scorpions*, G.A. Polis, ed., Stanford University Press, Stanford, CA, pp. 64-160.
- Sissom, W. D., and Fet, V., 2000, Family Diplocentridae Karsch, 1880, in: *Catalog of the Scorpions of the World* (1758-1998), V. Fet, W. D. Sissom, G. Lowe and M. E. Braunwalder, The New York Entomological Society, New York, pp. 329-354.
- Skead, C. J., 1973, Zoo-historical gazetteer, Ann. Cape Prov. Mus. 10:1-259.
- Smith, G. T., 1966, Observations on the life history of the scorpion *Urodacus abruptus* Pocock (Scorpionida), and an analysis of its home sites, *Aust. J. Zool.* 14:383-398.
- Soleglad, M. E., and Fet, V. 2003, High-level systematics and phylogeny of the extant scorpions (Scorpiones: Orthosterni), *Euscorpius* 11:1-175.
- Stockwell, S. A., 1985, A new species of *Heteronebo* from Jamaica (Scorpiones, Diplocentridae), *J. Arachnol.* 13:355-361.
- Stockwell, S. A., 1989, Revision of the phylogeny and higher classification of scorpions (Chelicerata), Ph.D. Thesis, University of California, Berkeley.
- Stuckenberg, B. R., 1962, The distribution of the montane palaeogenic element in the South African invertebrate fauna, *Ann. Cape Prov. Mus.* 2:190-205.
- Surveyor-General (Republic of South Africa), 1970, *Index of Orange Free State Farms*, Department of Public Works and Land Affairs, Pretoria.
- Surveyor-General (Republic of South Africa), 1988, Alphabetical list of farms in the Province of Transvaal, Department of Public Works and Land Affairs, Pretoria.
- SWALU, 1967, Swalurama Farms Directory, SWALU, Windhoek.
- Tikader, B. K., and Bastawade, D. B., 1983, Fauna of India, Vol 3., Scorpions, Zoological Survey of India, Calcutta.
- Vachon, M., 1973 [1974], Étude des caractères utilisés pour classer les familles et les genres de scorpions (Arachnides). 1. La trichobothriotaxie en arachnologie. Sigles trichobothriaux et types de trichobothriotaxie chez les scorpions, Bull. Mus. Natn. Hist. Nat., Paris (3) 140:857-958.
- van Zinderen Bakker, E. M., 1962, Botanical evidence for Quaternary Climates in Africa, *Ann. Cape Prov. Mus.* 2:16-31.
- van Zinderen Bakker, E. M., 1969, The "arid corridor" between south-western Africa and the Horn of Africa, *Palaeoecol. Afr.* 4:139-140.
- van Zinderen Bakker, E. M., 1975, The origin and palaeo-environment of the Namib Desert biome, *J. Biogeography* 2:65-73.
- van Zinderen Bakker, E. M., 1976, The evolution of late-Cenozoic palaeo-climates of southern Africa, *Palaeoecol. Afr.* 9:160-202.
- van Zinderen Bakker, E. M., 1978, Quaternary vegetation changes in southern Africa, in: *Biogeography and Ecology of southern Africa*, M. J. A. Werger, ed., Junk, The Hague, pp. 131-143.
- Warburg, M. R., and Ben-Horin, A., 1978, Temperature and humidity effects on scorpion distribution in northern Israel, *Symp. zool. Soc. Lond.* (London) **42**:161-169.
- Warburg, M. R., Goldenberg, S., and Ben-Horin, A., 1980, Scorpion species diversity and distribution within the Mediterranean and arid regions of northern Israel, *J. Arid Environ.* 3:205-213.
- Webb, R. S., 1950, Gazetteer for Basutoland. The first draft of a list of names, with special reference to the 1:250 000 map G. S. G. S. No. 2567 of June 1911: "Basutoland, from a reconnaissance survey made in 1904-9 by Captain M. C. Dobson, R. F. A.", A. H. Fischer and Sons, Paarl.

- Werger, M. J. A., 1978, Biogeography and Ecology of southern Africa, Junk, The Hague. Werner, F., 1936, Neu-Eingänge von Skorpionen im Zoologischen Museum in Hamburg, Festschr. 60. Geburtstage Prof. Dr. Embrik Strand 2:171-193.
- Williams, S. C., 1970, Coexistence of desert scorpions by differential habitat preference, Pan-Pacific Entomol. 46:254-267.
- Williams, S. C., 1980, Scorpions of Baja California, Mexico, and adjacent islands, *Occas. Pap. California Acad. Sci.* 135:1-127.
- Williams, S. C., 1987, Scorpion bionomics, Ann. Rev. Entomol. 32:275-295.

Appendix 1. Scorpion genera and species recorded from southern African countries, south of 15° latitude. Questionmarks reflect suspected occurrences that remain to be verified. Abbreviations as follows: A (Angola); B (Botswana); L (Lesotho); Ma (Malawi); Mo (Mozambique); N (Namibia); SA (South Africa); S (Swaziland); Za (Zambia); Zi (Zimbabwe); E (Extralimital).

| Species                                      | Α     | В  | L | Ma | Mo | N | SA | S | Za | Zi | Е |
|--|-------|----|---|----|----|---|----|---|----|----|---|
| Bothriuridae                                 | al 's | in |   |    |    |   |    |   |    |    |   |
| Brandbergia haringtoni Prendini, 2003        |       |    |   |    |    | + |    |   |    |    |   |
| Lisposoma elegans Lawrence, 1928             |       |    |   |    |    | + |    |   |    |    |   |
| Lisposoma joseehermanorum Lamoral, 1979      |       |    |   |    |    | + |    |   |    |    |   |
| Buthidae                                     |       |    |   |    |    |   |    |   |    |    |   |
| Afroisometrus minshullae (FitzPatrick, 1994) |       |    |   |    |    |   | +  |   |    | +  |   |
| Hottentotta arenaceus (Purcell, 1901)        |       |    |   |    |    | + | +  |   |    |    |   |
| Hottentotta conspersus (Thorell, 1876)       | +     |    |   |    |    | + |    |   |    |    |   |
| Hottentotta trilineatus (Peters, 1861)       |       | +  |   | ?  | +  |   | +  |   | +  | +  | + |
| Karasbergia methueni Hewitt, 1913            |       |    |   |    |    | + | +  |   |    |    |   |
| Lychas burdoi (Simon, 1882)                  |       |    |   | +  | +  |   | +  |   | +  | +  | + |
| Parabuthus brevimanus (Thorell, 1876)        | +     |    |   |    |    | + | +  |   |    |    |   |
| Parabuthus calvus Purcell, 1898              |       |    |   |    |    |   | +  |   |    |    |   |
| Parabuthus capensis (Ehrenberg, 1831)        |       |    |   |    |    | + | +  |   |    |    |   |
| Parabuthus distridor Lamoral, 1980           |       |    |   |    |    |   | +  |   |    |    |   |
| Parabuthus gracilis Lamoral, 1979            |       |    |   |    |    | + |    |   |    |    |   |
| Parabuthus granulatus (Ehrenberg, 1831)      | +     | +  |   |    |    | + | +  |   |    | +  |   |
| Parabuthus kalaharicus Lamoral, 1977         |       | +  |   |    |    | + | +  |   |    |    |   |
| Parabuthus kraepelini Werner, 1902           | ?     |    |   |    |    | + |    |   |    |    |   |
| Parabuthus kuanyamarum Monard, 1937          | +     | +  |   |    |    | + | +  |   | +  | +  |   |
| Parabuthus laevifrons (Simon, 1888)          |       | +  |   |    |    | + | +  |   |    |    |   |
| Parabuthus mossambicensis (Peters, 1861)     |       | +  |   | ?  | +  |   | +  |   | +  | +  |   |
| Parabuthus muelleri Prendini, 2000           |       |    |   |    |    | + |    |   |    |    |   |
| Parabuthus namibensis Lamoral, 1979          |       |    |   |    |    | + |    |   |    |    |   |
| Parabuthus nanus Lamoral, 1979               |       |    |   |    |    | + | +  |   |    |    |   |
| Parabuthus planicauda (Pocock, 1889)         |       |    |   |    |    |   | +  |   |    |    |   |
| Parabuthus raudus (Simon, 1888)              | +     | +  |   |    |    | + | +  |   | +  | +  |   |
| Parabuthus schlechteri Purcell, 1899         |       |    |   |    |    | + | +  |   |    |    |   |
| Parabuthus stridulus Hewitt, 1913            |       |    |   |    |    | + |    |   |    |    |   |
| Parabuthus transvaalicus Purcell, 1899       |       | +  |   |    | +  |   | +  |   |    | +  |   |
| Parabuthus villosus (Peters, 1862)           | ?     |    |   |    |    | + | +  |   |    |    |   |
| Pseudolychas ochraceus (Hirst, 1911)         |       | ?  | ? |    |    |   | +  |   |    | ?  |   |
| Pseudolychas pegleri (Purcell, 1901)         |       |    |   |    | +  |   | +  | + |    |    |   |
| Pseudolychas transvaalicus Lawrence, 1961    |       |    |   |    |    |   | +  |   |    |    |   |
| Uroplectes carinatus (Pocock, 1890)          | +     | +  |   |    |    | + | +  |   |    | +  |   |
| Uroplectes chubbi Hirst, 1911                |       | +  |   |    | +  |   | +  |   | +  | +  |   |
| Uroplectes flavoviridis Peters, 1861         |       |    |   | +  | +  |   | +  |   | +  | +  | ? |
| Uroplectes formosus Pocock, 1890             |       |    | + |    | +  |   | +  | + |    |    |   |
| Uroplectes gracilior Hewitt, 1914            |       |    |   |    |    | + | +  |   |    |    |   |
| Uroplectes insignis Pocock, 1890             |       |    |   |    |    |   | +  |   |    |    |   |
| Uroplectes lineatus (C. L. Koch, 1844)       |       |    |   |    |    |   | +  |   |    |    |   |
| Uroplectes longimanus Werner, 1936           |       |    |   |    |    | + |    |   |    |    |   |
| Uroplectes marlothi Purcell, 1901            |       |    |   |    |    |   | +  |   |    |    |   |
| Uroplectes olivaceus Pocock, 1896            |       |    |   |    | +  |   | +  | + |    | +  |   |
| Uroplectes of inhinguensis (Karsch, 1879)    | +     |    |   |    |    | + |    |   |    |    |   |

Appendix 1. (continued).

| Species                                   | Α | В | L | Ma     | Mo | N | SA      | S | Za | Zi      | Е   |
|---|---|---|---|--------|----|---|---------|---|----|---------|-----|
| Buthidae (continued)                      |   |   |   |        |    |   |         |   |    |         |     |
| Uroplectes planimanus (Karsch, 1879)      | + | + |   |        | +  | + | +       |   |    |         |     |
| Uroplectes schlechteri Purcell, 1901      |   |   |   |        |    |   | +       |   |    |         |     |
| Uroplectes teretipes Lawrence, 1966       |   |   |   |        |    |   |         |   |    |         |     |
| Uroplectes triangulifer (Thorell, 1876)   |   |   |   |        |    |   |         |   |    |         |     |
| Uroplectes tumidimanus Lamoral, 1979      |   | + |   |        |    |   |         |   |    |         |     |
| Uroplectes variegatus (C. L. Koch, 1844)  |   |   |   |        |    |   | ,       |   |    |         |     |
| Uroplectes vittatus (Thorell, 1876)       |   | + |   |        | +  | + | +       | + | +  | +       |     |
| Liochelidae                               |   |   |   |        |    |   |         |   |    |         |     |
| Cheloctonus anthracinus Pocock, 1899      |   |   |   |        |    |   | +       |   |    |         |     |
| Cheloctonus crassimanus (Pocock, 1896)    |   |   |   |        |    |   | +       |   |    |         |     |
| Cheloctonus glaber Kraepelin, 1896        |   |   |   |        |    |   | +       |   |    |         |     |
| Cheloctonus intermedius Hewitt, 1912      |   |   |   |        |    |   | +       |   |    |         |     |
| Cheloctonus jonesii Pocock, 1892          |   | ? |   |        | +  |   | +       | + | ?  | ?       |     |
| Hadogenes bicolor Purcell, 1899           |   |   |   |        |    |   | +       |   |    |         |     |
| Hadogenes gracilis Hewitt, 1909           |   |   |   |        |    |   | +       |   |    |         |     |
| Hadogenes granulatus Purcell, 1901        |   | + |   |        | +  |   |         |   | +  | +       |     |
| Hadogenes gunningi Purcell, 1899          |   |   |   |        |    |   | +       |   |    |         |     |
| Hadogenes lawrencei Newlands, 1972        |   |   |   |        |    |   |         |   |    |         |     |
| Hadogenes longimanus Prendini, 2001       |   |   |   |        |    |   | +       |   |    |         |     |
| Hadogenes minor Purcell, 1899             |   |   |   |        |    |   |         |   |    |         |     |
| Hadogenes newlandsi Prendini, 2001        |   |   |   |        |    |   |         |   |    |         |     |
| Hadogenes phyllodes Thorell, 1876         |   |   |   |        |    | + | +       |   |    |         |     |
| Hadogenes taeniurus (Thorell, 1876)       | + |   |   |        |    | + | tol to  |   |    |         |     |
| Hadogenes tityrus (Simon, 1888)           |   |   |   |        |    | + | +       |   |    |         |     |
| Hadogenes trichiurus (Gervais, 1843)      |   |   |   |        |    |   |         |   |    |         |     |
| Hadogenes troglodytes (Peters, 1861)      |   | + |   |        | +  |   | + 2     |   |    | +       |     |
| Hadogenes zuluanus Lawrence, 1937         |   |   |   |        |    |   |         |   |    |         |     |
| Hadogenes zumpti Newlands, 1985           |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus asper (Peters, 1861)       |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus basutus Lawrence, 1955     |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus capensis Thorell, 1876     |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus diremptus (Karsch, 1879)   |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus laevipes (Pocock, 1893)    |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus lamorali Lourenço, 1981    |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus piscatorius Lawrence, 1955 |   |   |   |        |    |   |         |   |    |         |     |
| Opisthacanthus rugiceps Pocock, 1897      |   |   |   | +      |    |   |         |   |    |         |     |
| Opisthacanthus rugulosus Pocock, 1896     |   |   |   | +      |    |   |         |   |    |         |     |
| Opisthacanthus validus Thorell, 1876      |   |   | + |        |    |   |         |   |    |         |     |
| Scorpionidae                              |   |   |   |        |    |   |         |   |    |         |     |
| Opistophthalmus adustus Kraepelin, 1908   |   |   |   |        |    |   |         |   |    |         |     |
| Opistophthalmus ammopus Lamoral, 1980     |   |   |   |        |    |   |         |   |    |         |     |
| Opistophthalmus ater Purcell, 1898        |   |   |   |        |    |   |         |   |    |         |     |
| Opistophthalmus austerus Karsch, 1879     |   |   |   |        |    |   |         |   |    |         |     |
| Opistophthalmus boehmi (Kraepelin, 1896)  |   |   |   |        |    |   |         |   |    |         | 100 |
|   |   | + |   |        |    |   |         |   |    |         | +   |
| Opistophthalmus brevicauda Lawrence, 1928 |   |   |   |        |    |   |         |   |    |         |     |
| Opistophthalmus capensis (Herbst, 1800)   |   | , |   |        |    |   |         |   |    |         |     |
| Opistophthalmus carinatus (Peters, 1861)  | + | + |   |        |    |   |         |   |    |         |     |
| Opistophthalmus cavimanus Lawrence, 1928  |   |   |   | Digiro |    | + | 811 (0) |   |    | 13 E.13 | 1.7 |

Appendix 1. (continued).

| Species                                     | Α | В | L | Ma | Mo | N | SA   | S    | Za   | Zi   | E    |
|---|---|---|---|----|----|---|------|------|------|------|------|
| Scorpionidae (continued)                    |   |   |   |    |    |   | ibou | niin | 0015 | Shir | ro H |
| Opistophthalmus chrysites Lawrence, 1967    |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus coetzeei Lamoral, 1979      |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus concinnus Newlands, 1972    |   | + |   |    |    | + | +    |      |      |      |      |
| Opistophthalmus crassimanus Purcell, 1899   |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus fitzsimonsi Hewitt, 1935    |   | + |   |    |    |   |      |      |      |      |      |
| Opistophthalmus flavescens Purcell, 1898    |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus fossor Purcell, 1898        |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus fuscipes Purcell, 1898      |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus gibbericauda Lamoral, 1979  | + |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus gigas Purcell, 1898         |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus glabrifrons Peters, 1861    |   | + |   | +: | +  |   | +    | ?    |      | +    | +    |
| Opistophthalmus granicauda Purcell, 1898    |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus granifrons Pocock, 1896     |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus haackei Lawrence, 1966      |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus harpei Harington, 2001      |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus holmi (Lawrence, 1969)      |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus intercedens Kraepelin, 1908 |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus intermedius Kraepelin, 1894 |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus jenseni (Lamoral, 1972)     |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus karrooensis Purcell, 1898   |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus keilandsi Hewitt, 1914      |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus lamorali Prendini, 2000     |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus laticauda Purcell, 1898     |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus latimanus C. L. Koch, 1841  |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus latro Thorell, 1876         |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus lawrencei Newlands, 1969    |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus leipoldti Purcell, 1898     |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus litoralis Lawrence, 1955    | + |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus longicauda Purcell, 1899    |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus lornae Lamoral, 1979        |   |   |   |    |    | + | +    |      |      |      |      |
| Opistophthalmus luciranus Lawrence, 1959    | + |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus macer Thorell, 1876         |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus nitidiceps Pocock, 1896     |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus opinatus (Simon, 1888)      |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus pallipes C. L. Koch, 1842   |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus pattisoni Purcell, 1899     |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus penrithorum Lamoral, 1979   |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus peringueyi Purcell, 1898    |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus pictus Kraepelin, 1894      |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus pluridens Hewitt, 1918      |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus praedo Thorell, 1876        |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus pugnax Thorell, 1876        |   |   | + |    |    |   | +    |      |      |      |      |
| Opistophthalmus pygmaeus Lamoral, 1979      |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus scabrifrons Hewitt, 1918    |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus schlechteri Purcell, 1898   |   |   |   |    |    |   | +    |      |      |      |      |
| Opistophthalmus schultzei Kraepelin, 1908   |   |   |   |    |    |   |      |      |      |      |      |
| Opistophthalmus setifrons Lawrence, 1961    |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus ugabensis Hewitt, 1934      |   |   |   |    |    | + |      |      |      |      |      |
| Opistophthalmus wahlbergii (Thorell, 1876)  | + | + |   |    |    | + | +    |      | +    | +    |      |